

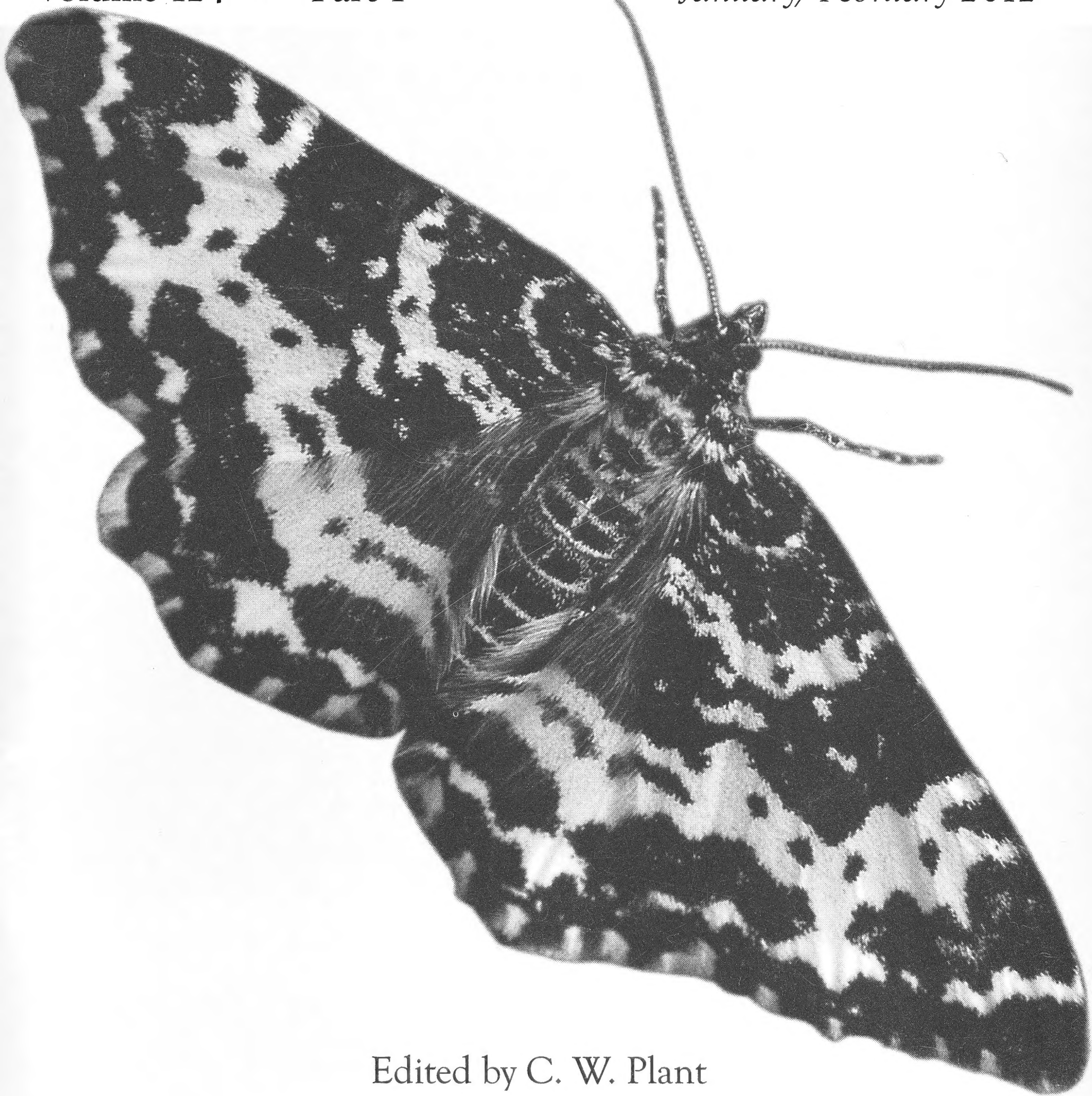
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Editor

C.W. PLANT, B.Sc., F.R.E.S.

14 West Road, Bishops Stortford, Hertfordshire CM23 3QP.

Telephone: 01279 507697 E-mail: cpauk1@ntlworld.com

Who to contact

Material for publication, books for review, obituaries, advertisements – the Editor
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Please follow the format in this issue. The Editor will be pleased to assist in any way possible, especially if you are a first-time author. If in doubt – contact me! Ideally material will be compatible with Microsoft Word and will arrive via e-mail, but we will accept typescripts or *neatly* hand-written contributions for shorter Notes.

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County Moth Recorders and Moth Recording Schemes

Formal moth recorders for the vice-counties of the British Isles and the Channel Islands have, in many cases, existed since the late Victorian period and were traditionally appointed by the county field club or natural history society. At the start of 2012, around 30 vice-counties continue to be covered by such formally appointed Recorders. Remaining counties are, however, not neglected and in recent years many gaps in this traditional system have been filled through the agency of *Butterfly Conservation* and the *Moths Count* project. Together, these County Moth Recorders (CMRs) are the people to whom all moth records should be sent in order to ensure that a permanent record is maintained; for this purpose a list of formal CMRs is printed below.

Systems are now in place for direct and rapid communication between all of the CMRs and there is also full two-way communication between CMRs and the various taxon-based recording schemes (including the National Moth Recording Scheme); these are also listed below. If you do not already submit moth records to the appropriate CMR please do consider doing so in the future.

In the following list, the named CMR records both macros and micros unless otherwise stated:

VC1 & VC 2 Cornwall

Tony James, Manzil, Trewithen Terrace, Greatwork, Nr Ashton, Helston, Cornwall, TR13 9TQ. (E-mail: cmg.com).ms@btinternet.com).

VC3 & VC4 South Devon & North Devon

Roy McCormick, 36 Paradise Road, Teignmouth, Devon, TQ14 8NR. (E-mail: 181525131225.devmg@talktalk.net).

VC5 & VC6 South Somerset and North Somerset

Macros: John Bebbington FRPS, Quantock View, Newtown, Langport, TA10 9SE. (E-mail: john.bebbingtonfrps@btinternet.com).

Micros: Vacant.

VC7 & VC8 North Wiltshire & South Wiltshire

Macros: Marc S Taylor, 12 Durham Street, Swindon, Wiltshire, SN1 3HS. (E-mail: wilts_moth_group@yahoo.co.uk).

Micros: Godfrey & Michael Smith, Bullen Hill Farm, Steeple Ashton Common, Trowbridge, Wiltshire, BA14 6DY.

VC9 Dorset

Macros: Les Hill, 48 East Lodge, East Lulworth, Wareham, Dorset, BH20 5QL. (E-mail: les@mklep.co.uk).

Micros: Phil Sterling, Natural Environment Manager, Environment Services, Dorset County Council, County Hall, Dorchester, Dorset, DT1 1XJ. (E-mail: p.sterling@dorsetcc.gov.uk).

VC10 Isle of Wight

Sam Knill-Jones, 1 Moorside, Moons Hill, Totland, Isle of Wight, PO39 OHU.

VC11 & VC12 South Hampshire & North Hampshire

Macros: Tim Norriss, 40 Taskers Drive, Anna Valley, Andover, Hampshire, SP11 7SA. (E-mail: tim@kitsmail.com).

Micros: Mike Wall, 11 Waterloo Avenue, Basingstoke, Hampshire, RG23 8DL. (E-mail: mike@hantsmoths.org.uk).

VC13 & VC14 West Sussex & East Sussex

Colin Pratt, 5 View Road, Peacehaven, East Sussex, BN10 8DE. (E-mail: colin.pratt@talk21.com).

VC15 & VC16 East Kent & West Kent

Macros: Ian Ferguson, 31 Cathcart Drive, Orpington, Kent, BR6 8BU. (E-mail: ianD.ferguson@hemscott.net).

Micros: David Agassiz, The Garden House, Stafford Place, Weston-super-Mare, BS23 2QZ. (E-mail: agassiz@btinternet.com).

VC17 Surrey

Macros: Graham Collins, 15 Hurst Way, South Croydon, Surrey, CR2 7AP. (E-mail: stigmella.aurella@tiscali.co.uk).

Micros: Jim Porter, 4 Orchard Road, Chessington, Surrey, KT9 1AN. (E-mail: jimoth17@hotmail.co.uk).

VC18 & VC19 South Essex & North Essex

Macros: Vacant.

Micros: Stephen Rolls, 11 Caxton Close, Tiptree, Essex, CO5 0HA. (E-mail: bunnrolls@aol.com).

Records collator: Peter R. Harvey BSc, 32 Lodge Lane, Grays, Essex RM16 2YP. (E-mail: grays@peterharvey.freemove.co.uk).

VC20 & VC21 Hertfordshire & Middlesex

Colin Plant, 14 West Road, Bishops Stortford, Hertfordshire, CM23 3QP. (E-mail: cpauk1@ntlworld.com).

VC22 Berkshire

Martin Harvey, Evermor, Bridge Street, Great Kimble, Aylesbury, Buckinghamshire, HP17 9TN. (E-mail: kitenetter@gmail.com).

VC23 Oxfordshire

Macros: Martin Townsend, 26 Bartholomew Road, Cowley, Oxford, OX4 3QQ. (E-mail: martin.townsend4@ntlworld.com).

Micros: Martin Corley, Pucketty Farm Cottage, Faringdon, Oxfordshire, SN7 8JP. (E-mail: martin.corley@btinternet.com).

VC24 Buckinghamshire

Martin Albertini, 11 Hargrave Road, Maidenhead, Berkshire, SL6 6JR. (E-mail: malbertini@onetel.com).

VC25 & VC26 East Suffolk & West Suffolk

Tony Prichard, 3 Powling Road, Ipswich, Suffolk, IP3 9JR. (E-mail: tony@suffolkmoths.org.uk).

VC27 & VC28 East Norfolk & West Norfolk

Jim Wheeler, Iveygreen, Town St, Upwell, Norfolk, PE14 9AD. (E-mail: jim@norfolkmoths.org.uk).

VC29 Cambridgeshire

John Dawson, Lodge Farm, Fulbourn, Cambridge, CB1 5HR. (E-mail: jd.vc29@btinternet.com).

VC30 Bedfordshire

Macros: Andy & Melissa Banthorpe, 32 Long Close, Lower Stondon, Bedfordshire, SG16 6JS. (E-mail: macromoths@bnhs.org.uk).

Micros: David Manning, 27 Glebe Rise, Sharnbrook, Bedford, MK44 1JB. (E-mail: micromoths@bnhs.org.uk).

VC31 Huntingdonshire

Barry Dickerson, 27 Andrew Road, Eynesbury, St Neots, Cambridgeshire, PE19 2QE. (E-mail: barry@eynesbury27.freemove.co.uk).

VC32 Northamptonshire

Macros: John Ward, 109 Blandford Avenue, Kettering, Northamptonshire, NN16 9AS. (E-mail: bjward@btinternet.com).

Micros: David Manning, 27 Glebe Rise, Sharnbrook, Bedford, MK44 1JB. (E-mail: micromoths@bnhs.org.uk).

VC33 & VC34 East Gloucestershire & West Gloucestershire

Roger Gaunt, Firtree Cottage, Cold Harbour, St Briavels, Lydney, Gloucestershire, GL15 6SB. (E-mail: roger.gaunt@btinternet.com).

VC35 Monmouthshire

Macros: Martin Anthoney, 23 Malvern Close, Risca, Newport, Gwent, NP11 6QY. (E-mail: martin@chemlep.demon.co.uk).

Micros: Sam Bosanquet, Cnwc-y-llwyn, Brechfa, Carmarthen, SA32 7QR. (E-mail: s.bosanquet@ccw.gov.uk).

VC36 Herefordshire

Verification: Michael Harper, The Cherry Orchard, Gloucester Road, Bullen, Ledbury, Herefordshire, HR8 2JE.

Record Collator: Phyl King, Woodlea, Little Dew Church, Hereford, HR2 6PS. (E-mail: phyl.woodlea@gmail.com).

VC37 Worcestershire

Tony Simpson, The Sycamores, Old Rectory Gardens, Leigh, Worcestershire, WR6 5LD. (E-mail: tonysimpson.1945@hotmail.co.uk).

VC38 Warwickshire

Macros: David Brown, Jacksons Lawn, Charlecote, Nr. Warwick, CV35 9EW. (E-mail: davidcgbrown@yahoo.co.uk).

Micros: Nigel Stone, 8 Grove Lane, Keresley End, Coventry, CV7 8PN. (E-mail: njstone@hotmail.com).

VC39 Staffordshire

David Emley, Earth Sciences & Geography, Keele University, Staffordshire, ST5 5BG. (E-mail: d.w.emley@esci.keele.ac.uk).

VC40 Shropshire

Macros: Tony Jacques, 49 Foundry Lane, Broseley, Shropshire, TF12 5DN. (E-mail: vc40@talktalk.net).

Micros: Godfrey Blunt, 8 Holmes Orchard, Alveley, Bridgnorth, Shropshire, WV15 6NX. (E-mail: arthur.blunt195@btinternet.com).

VC41 Glamorgan

Dave Slade, South East Wales Biodiversity Records Centre, 13 St Andrews Crescent, Cardiff, CF10 3DB. (E-mail: david.slade@sewbrec.org.uk).

VC42 Breconshire

Norman Lowe, 6 Tai Canol, Llangorse, Brecon, Powys, LD3 7UR. (E-mail: norman@enviro-consulting.com).

VC43 Radnorshire

Pete & Ginny Clarke, 9 Dany-bryn, Glasbury on Wye, Hereford, HR3 5NH. (E-mail: peteandginnyc@tiscali.co.uk).

VC44 Carmarthenshire

Jon Baker, 14 Job's Well Road, Carmarthen, SA31 3HG. (E-mail: Mothboy@btinternet.com).

VC45 Pembrokeshire

Paul Warren, 39 Pill Road, Milford Haven, Pembrokeshire, SA73 2NS. (E-mail: paul.warren@rocketmail.com).

VC46 Cardiganshire

Peter Walters Davies, Alltgoy, Caemelyn, Aberystwyth, Ceredigion, SY23 2HA. (E-mail: davies1912@tiscali.co.uk).

VC47 Montgomeryshire

Peter Williams, Pandy, Commings Coch, Machynlleth, Powys, SY20 8LG. (E-mail: peterwilliams526@btinternet.com).

VC48 Merionethshire

Andrew Graham, c/o Trawscoed, Llanuwchllyn, Bala, Gwynedd, LL23 7TD. (E-mail: angrhm@globalnet.co.uk).

VC49 Caernarvonshire

Julian Thompson, Pensychnant Conservation Centre, Sychnant Pass, Conwy, LL32 8BJ. (E-mail: julian.pensychnant@btinternet.com).

VC50 Denbighshire

Macros: Justin Williams, The Old Orchard, Sunnyridge Avenue, Marford, LL12 8TE. (E-mail: northwalesmoths@hotmail.co.uk).

Micros: Bryan Formstone, 15 Beech Avenue, Gresford, Wrexham, LL12 8EL. (E-mail: bryanformstone@dsl.pipex.com).

VC51 Flintshire

macros: Justin Williams, The Old Orchard, Sunnyridge Avenue, Marford, LL12 8TE. (E-mail: northwalesmoths@hotmail.co.uk).

micros: vacant.

VC52 Anglesey

John Harold, Hen Ardd, Carreg y Gath, Rhiwlas, Bangor, LL57 4HD. (E-mail: jhmoths@yahoo.co.uk).

VC53 & VC54 South Lincolnshire & North Lincolnshire

Colin Smith, 3 Dear Street, Market Rasen, Lincolnshire, LN8 3BH. (E-mail: csmith668@btinternet.com).

VC55 Leicestershire (with Rutland)

Adrian Russell, 15 St Swithins Road, Leicester, LE5 2GE. (E-mail: adrian@wainscot.demon.co.uk).

VC56 Nottinghamshire

Macros: Sheila Wright, Nottingham Natural History Museum, Wollaton Hall, Wollaton Park, Nottingham, NG8 2AE. (E-mail: sheila.wright@nottinghamcity.gov.uk).

Micros: Dave Budworth, 121 Wood Lane, Newhall, Swadlincote, Derbyshire, DE11 0LX. (E-mail: dbud01@aol.com).

VC57 Derbyshire

Dave Budworth, 121 Wood Lane, Newhall, Swadlincote, Derbyshire, DE11 0LX. (E-mail: dbud01@aol.com).

VC58 Cheshire

Macros: Vacant

Micros: Stephen Hind, 32 Carleton Road, Higher Poynton, Stockport, Cheshire, SK12 1TL. (E-mail: stevehind32@btinternet.com).

VC59 & VC60 South Lancashire & West Lancashire

Macros: Graham Jones, 45 Liverpool Old Road, Walmer Bridge, Preston, Lancashire, PR4 5QA. (E-mail: gra_ham_jones@yahoo.co.uk).

Micros: Stephen Palmer, 137 Lightfoot Lane, Fulwood, Preston, Lancashire, PR4 0AH. (E-mail: s.palmer12@btopenworld.com).

VC61, VC62, VC63, VC64 & VC65 South-east Yorkshire, North-east Yorkshire, South-west Yorkshire, Mid-west Yorkshire, North-west Yorkshire

Macros: Charles Fletcher, The Forge, Hutton Conyers, Ripon, N Yorks, HG4 5EB. (E-mail: chfletcher@btinternet.com).

Micros: Harry Beaumont, 37 Melton Green, West Melton, Rotherham, South Yorkshire, S63 6AA.

Records should be sent to the following vice-county coordinators:

VC61 South-east Yorkshire

Tony Ezard, Greenacre Cottage, Eastgate, Rudston, YO25 4UX. (E-mail: anthony.ezard@homecall.co.uk).

VC62 North-east Yorkshire

David Chesmore, 39 Hawthorn Drive, Holme upon Spalding Moor, York, YO43 4HX. (E-mail: edc1@ohm.york.ac.uk).

VC63 South-west Yorkshire

Mike Couling, High Up, Binns Lane, Holmfirth, HD9 3BJ. (E-mail: mike_couling@yahoo.co.uk).

VC64 & 65 Mid-west Yorkshire & North-west Yorkshire

Charles Fletcher, The Forge, Hutton Conyers, Ripon, N Yorks, HG4 5EB. (E-mail: chfletcher@btinternet.com).

VC66 Durham

Macros: Keith Dover, 4 Lindisfarne Avenue, Chester-le-Street, Co Durham, DH3 3PT. (E-mail: k.dover879@btinternet.com).

Micros: Tim Barker, Tap and Spile, 27 Front Street, Framwellgate Moor, Durham, DH1 5EE. (E-mail: tim@tapandspile.co.uk).

VC67 & VC68 South Northumberland & North Northumberland

Tom Tams. (E-mail: tomsphotos@hotmail.co.uk).

VC69 & VC70 Westmorland with North Lancashire & Cumberland

Record Collator: Teresa Frost, Cumbria Biodiversity Data Centre, Tullie House Museum, Castle Street, Carlisle CA3 8TP. (E-mail: manager@cbdc.org.uk).

VC71 Isle of Man

Ian Scott. (E-mail: scotty1@manx.net).

VC72, VC73 & VC74 Dumfries-shire, Kirkcudbrightshire & Wigtownshire

Keith Naylor, c/o Dumfries and Galloway Environmental Resources Centre, Solway Heritage, 7 Church Crescent, Dumfries, DG1 1DF. (E-mail: mothrecorder@dgerc.org.uk).

VC75 & VC76 Ayrshire, Inverclyde & Renfrewshire

Neil Gregory, 32 Oldhall Drive, Kilmacolm, Inverclyde, PA13 4RF. (E-mail: droitwich@btinternet.com).

VC 77 Lanarkshire

Richard Weddle, 89 Novar Drive, Glasgow, G12 9SS. (E-mail: richard.weddle@tiscali.co.uk).

VC78 Peebleshire

Reuben Singleton, 5 Frankscroft, Peebles, EH45 9DX. (E-mail: reuben@dukehaugh.free-online.co.uk).

VC 79 Selkirkshire

Malcolm Lindsay, Burn House, Mossilee Road, Galashiels, TD1 1NF. (E-mail: malcandles@tiscali.co.uk).

VC80 Roxburghshire

Andy Fitchett, 1 The Croft, Nether Blainslie, Galashiels, TD1 2 QF. (E-mail: andy.fitchett@virgin.net).

and Jeff Waddell, 33 Eildon View, Melrose, Roxburghshire, TD6 9RH. (E-mail: jeffwaddell11@yahoo.co.uk).

VC81 Berwickshire

Barry Prater, Fortview, 12 Barefoots Crescent, Eyemouth, Berwickshire, TD14 5BA. (E-mail: barry@prater.myzen.co.uk).

VC82 East Lothian

Mike Thornton, 2/4 Piershill Terrace, Edinburgh, EH8 7ES. (E-mail: mike_thornton_99@yahoo.com).

VC83 Midlothian

Alastair Sommerville, 4 Woodbank Crofts, Westfield, Bathgate, West Lothian, EH48 3AT. (E-mail: alastair@biodiversitysolutions.co.uk).

VC84 West Lothian

Mark Cubitt, 12 Burgh Mills Lane, Linlithgow, EH49 7TA. (E-mail: moths@cubes.f9.co.uk).

VC 85 Fife

Duncan Davidson, 140 Pitcorthie Drive, Dunfermline, KY11 8BJ. (E-mail: Duncan@dwwd.freemove.co.uk).

VC86 & VC87 Stirlingshire, West Perthshire

John Knowler, 3 Balfleurs Street, Milngavie, Glasgow, G62 8HW. (E-mail: john.knowler@ntlworld.com).

VC88 Mid Perthshire

Julie Stoneman. (E-mail: jstoneman@gmail.com).

VC89 East Perthshire

John Thorpe, Pipers Croft, Killiecrankie, Perthshire, PH16 5LW. (E-mail: johnethorpe@btinternet.com).

VC 90 Angus

David Lampard, 51 Johnston Avenue, Dundee DD3 8HA. (E-mail: dlampard@clara.co.uk).

VC91, VC92 & VC 93 Kincardineshire, South Aberdeenshire, North Aberdeenshire

Mark Young, Meiklepark, Oldmeldrum, AB51 0DL. (E-mail: m.young@abdn.ac.uk).

VC94 Banffshire

Roy Leverton, Whitewells, Ordiquhill, Cornhill, Banffshire, AB45 2HS. (E-mail: roy345@btinternet.com).

VC95 Moray

Mike Taylor, Allasdale, Kinchurdy Road, Boat of Garten, Inverness-shire, PH24 3BP. (E-mail: mst.taylor@btinternet.com).

VC96 East Inverness-shire

Pete Moore, Ivy Cottage, Insh, Kingussie, Inverness-shire, PH21 1NT. (E-mail: pete.moore@rspb.org.uk).

VC97 West Inverness-shire

Tom Prescott, Mill House, Mill Road, Kingussie, Inverness-shire, PH21 1LF. (E-mail: tomprescott@btconnect.com).

VC98 Argyll Main

Andrew Masterman, Flat 1/3, 214 Calder Street, Glasgow, G42 7PE. (E-mail: andrewmasterman@hotmail.com).

VC99 Dunbartonshire

John Knowler, 3 Balfleurs Street, Milngavie, Glasgow, G62 8HW. (E-mail: john.knowler@ntlworld.com).

VC100 Clyde Isles**Arran**

Kate Sampson, National Trust for Scotland Ranger Service, Brodick Country Park, Isle of Arran, KA27 8HY. (E-mail: ksampson@nts.org.uk).

Bute

Glyn Collis, Seasgair, Ascog, Isle of Bute, PA20 9ET. (E-mail: glyn@butemuseum.org).

Cumbraes

Neil Gregory, 32 Oldhall Drive, Kilmacolm, Inverclyde, PA13 4RF. (E-mail: droitwich@btinternet.com).

VC101 Kintyre

Helen Bibby, Gowanlea, Kilduskland Road, Ardrishaig, Argyll, PA30 8EH. (E-mail: helen.bibby@live.co.uk).

VC102 South Ebudes

John Armitage, Airigh Sgallaidh, Portnahaven, Isle of Islay, Argyll, PA47 7SZ. (E-mail: jsa@ornquest.plus.com).

VC 103 Mid Ebudes

Alan Skeates, Dunellen, Craignure, Isle of Mull, Argyll, PA65 6AY. (E-mail: alanskeates@btinternet.com).

VC104 North Ebudes**Skye**

Brian Neath, Culag, Carr Brae, Dornie, Kyle, Ross-shire, IV40 8HA. (E-mail: bandj@culag.plus.com).

Small Isles

Jessie Wormell, Fionchra, Carsfad, St John's Town of Dalry, Castle Douglas, DG7 3SU. (E-mail: entomology@carsfadcottagebandb.co.uk).

VC105 West Ross

Brian Neath, Culag, Carr Brae, Dornie, Kyle, Ross-shire, IV40 8HA. (E-mail: bandj@culag.plus.com).

VC106 East Ross

Margaret Currie and Andrew Scott, 7 Balnatua, Culbokie, Dingwall, Ross-shire, IV7 8JQ. (E-mail: moths.vc106@btinternet.com).

VC107 East Sutherland

Tony Mainwood, 13 Ben Bhraggie Drive, Golspie, Sutherland, KW10 6SX. (E-mail: tony.mainwood@btinternet.com).

VC108 West Sutherland

Sue Agnew, 78 Strathkanaird, Ullapool, Ross-shire, IV26 2TP. (E-mail: sue.agnew@virgin.net).

VC109 Caithness

Neil Money, Heathfield House, Dunnet, Thurso, Caithness, KW14 8XP. (E-mail: neil@heathfield99.plus.com).

VC110 Outer Hebrides

Steve Duffield, 5 Drimsdale, South Uist, Western Isles, HS8 5RT. (E-mail: steveduffield@hebrides.net).

VC111 Orkney Islands

Sydney Gauld, Quoyberstane, Work Road, St Ola, Kirkwall, Orkney, KW15 1UF. (E-mail: orklander45@gmail.com).

VC112 Shetland Islands

Shetland Biological Record Centre, Shetland Amenity Trust, Garthspool, Lerwick, ZE1 0NY. (E-mail: sbrc@shetlandamenity.org).

Fair Isle

Nick Riddiford, Schoolton, Fair Isle, Shetland, ZE2 9JU. (E-mail: taib@mallorcaweb.net).

VC113 Channel Islands**Alderney**

David Wedd, 2 La Vallee Clos, Alderney, Channel Islands, GY9 3XA. (E-mail: dwedd@cwgsy.net).

Guernsey

Rich Austin, Maymyo, Les Amballes, St Peter Port, Guernsey, GY1 1WY. (E-mail: raustin@guernsey.net).

Jersey and other Islands

Roger and Margaret Long, Ozarda, Les Hamonnets, St John, Jersey, JE3 4FP. (E-mail: randmlong@localdial.com).

Northern Ireland

Ted Rolston, 42 Beech Dene Gardens, Lisburn, BT28 3JH. (E-mail: ted_rolston@hotmail.com).

Republic of Ireland

Angus Tyner, Honeyoak, Cronykeery, Ashford, Co. Wicklow, Ireland. (E-mail: records@mothsireland.com).

National moth recording scheme addresses**National Macro Moth Recording Scheme**

Manor Yard, East Lulworth, Dorset BH20 5QP.

Crambidae and Pyralidae Recording Scheme

Tony Davis, 17 Orchards Way, West End Village, Southampton, SO30 3FB. (E-mail: pyralid.scheme@virgin.net).

Pterophoridae

Colin Hart, Fourpenny Cottage, Dungates Lane, Buckland, Betchworth, Surrey, RH3 7BD. (E-mail: colin@dungates.co.uk).

National Leaf-mining moth Recording Scheme

Rob Edmunds, Embankment House, Farthing Road, Downham Market, Norfolk, PE38 0AF. (E-mail: rdedmunds@btinternet.com).

Incurvarioidea Recording Scheme (Longhorn moths and allies)

Keith Bland, 35 Charterhall Road, Edinburgh, EH9 3HS.

Gelechiidae Recording Scheme

Carolyn and Stephen Palmer, 137 Lightfoot Lane, Fulwood, Preston, Lancashire PR4 0AH. (E-mail: s.palmer12@btopenworld.com).

National Review of Microlepidoptera

John Langmaid, Wilverley, 1 Dorrita Close, Southsea, Hampshire PO4 0NY (E-mail: johnrendle.langmaid@gmail.com) and Mark Young, Meiklepark, Oldmeldrum, Aberdeenshire AB51 ODL. (E-mail: m.young@abdn.ac.uk).

National Migrant Lepidoptera Recorder

Sean Clancy, 1 Myrtle Villas, Sussex Road, New Romney, Kent TN28 8DY. (E-mail: trapsite@talktalk.net).

***Monochroa lutulentella* (Zeller) (Lep.: Gelechiidae) new to the Isle of Wight, with details of records from Hampshire**

On 31 May 2011 the authors led a moth recording and conservation workshop for the Hampshire and Isle of Wight Branch of Butterfly Conservation. This included running moth traps in the evening, at Cranmore SSSI, near Yarmouth, on the Isle of Wight (O. S. grid reference SZ 396903).

MCH was running a 125 watt mercury vapour bulb over a white sheet, and this was switched on at dusk. Almost immediately an unfamiliar gelechiid moth started to arrive in numbers on the sheet, with at least 12 seen in total. Further individuals were attracted to a nearby Skinner trap run by TN on the edge of the footpath leading through this part of the SSSI.

Three specimens were kept, which upon close examination were identified by MCH as *Monochroa lutulentella*. This has two colour forms, one with very dark brown forewings and the other with paler brown forewings, both of which were present. Two male specimens were subsequently dissected, and the identification was confirmed by John Langmaid from photos of the moths and dissected genitalia. The specimens have a wingspan of 17.5mm, larger than the range of 14-16mm given in Emmet and Langmaid ([Eds.] 2002. *The Moths and Butterflies of Great Britain and Ireland* 4(2). Harley Books), but online sources give a wingspan of up to 18mm for this species (e.g. www2.nrm.se/en/svenska_fjarilar/m/monochroa_lutulentella.html).

Monochroa lutulentella has been recorded quite widely in south-east England (Emmet and Langmaid *op. cit.*), but is clearly very local, and has been proposed for Red Data Book (Insufficiently Known) status (Parsons. 1995. *UK Nature Conservation* 16. JNCC). The database for Hampshire and Isle of Wight micro-moth records, maintained by Mike Wall, has no previous record for the Isle of Wight (VC 10), and in fact the only record on the database for modern Hampshire falls in VC 8, South Wiltshire (Lower Breach Copse, West Park, July 2002, recorded by David G. Green). However, the map in Emmet and Langmaid *op. cit.* also shows a record for South Hampshire (VC 11), based on nine specimens in the National Museums Collection Centre, Edinburgh. All of them are in the B. W. Weddell collection (accession details: B.W.Weddell / RSM 1976.016), and all are labelled 'New Forest'. Eight of them were collected by Weddell: three examples dated 1.vii.1929, three dated 3.vii.1929, one dated 14.vi.1934 and one dated 16.vi.1934. The ninth specimen has the data 'New Forest / 1.7.1929 / A. R. Hayward', and was probably taken by Hayward while collecting with Weddell (R. Lyszkowski *pers. comm.*).

Records for *Monochroa lutulentella* are from a wide range of habitats; it is associated with wetland habitats (e.g., there are recent records at Wicken Fen, J.R. Langmaid *pers. comm.*), but has also been found in dry fields and gardens (Emmet and Langmaid *op. cit.*). Cranmore SSSI is notable for its acid grass/heathland habitats. These don't match well to the National Vegetation Classification plant communities, but do have affinities with similar habitats in the New Forest (Cox,

2001. *Note on Cranmore grass heathland vegetation, as briefing for SSSI notification*. Unpublished report to English Nature). Cranmore SSSI also contains areas of scrub and mixed woodland. The moth trap was sited in a relatively small area of grass/heath, with a conifer plantation on one side and mixed broadleaf scrub and hedgerow on other sides. The foodplant for *Monochroa lutulentella* is not confirmed in the UK. Larvae are reported to feed on the roots of Meadowsweet *Filipendula ulmaria* on the continent of Europe (Emmet and Langmaid *op. cit.*), but this plant has not been recorded on Cranmore SSSI.

Thanks to Dr J. R. Langmaid for confirming the identification and supplying data from the *MBGBI* maps originally compiled by Maitland Emmet, to Richard Lyszkowski and Dr Keith Bland for providing information on the specimens in the National Museums Collection Centre, Edinburgh, to Mike Wall for checking the Hampshire and Isle of Wight moth database and to the Hampshire and Isle of Wight Branch of Butterfly Conservation for funding the moth workshop. — MARTIN C. HARVEY, Evermore, Bridge Street, Great Kimble, Aylesbury HP17 9TN (E-mail: kitenetter@googlemail.com) & TIM NORRISS, 40 Taskers Drive, Anna Valley, Andover, Hampshire SP11 7SA (E-mail: tim@kitsmail.com).

Results from a pheromone trap run in a rural Norfolk garden during 2011

A single pheromone lure and trap intended for *Grapholita funebrana* (Treitschke, 1835) was deployed in my in-laws garden at Cawston in rural Norfolk from the 1st May to the 22nd September 2011. The purpose was to help reduce this species which is a constant pest of their plum trees.

The same lure was left *in situ* with the trap that was filled with water to a desired level to contain and destroy the moths during this period. Approximately every month I emptied the contents into Isopropyl Alcohol to preserve and work with at a later date.

This I was able to do after the Christmas break and to my surprise, amongst 66 specimens dissected, 58 were of the intended species but a further seven were *Pammene albuginana* (Guenée, 1845) constituting the first records for Norfolk. In addition a single *Grapholita tenebrosana* (Duponchel, 1843) was also determined, the first Norfolk record since 1988.

This was not totally unexpected though as it is widely known that synthetic pheromone lures do have the potential to draw in more than the intended species.

Many thanks to Jim Wheeler for providing the relevant species data. — JON CLIFTON, Kestrel Cottage, Station Road, Hindolveston, Norfolk.

One to watch out for: *Tuta absoluta* (Meyrick) (Lep.: Gelechiidae) apparently free-flying in Worcestershire

An invitation to moth trap at Windmill Hill near Evesham, Worcestershire in July 2010 led to an interesting encounter with the tomato-loving South American micro moth *Tuta absoluta*, which first appeared on this side of the Atlantic in 2006 when it was discovered in Spain (see <http://www.tutaabsoluta.com>). It has since been sweeping across Europe and North Africa, apparently causing serious damage to tomato crops. It reached Britain at Cheltenham, Gloucestershire in 2009 (Homan, 2010. *Ent. Rec.* **122**: 31-32) and was also noted in Yorkshire (Langmaid, 2010. *Ent. Rec.* **122**: 245). It was first recorded in Wales in Caernarvonshire in 2010 (Langmaid, *Ent. Rec.*, **123**: 265). Since then a number of “outbreaks” have been reported to the Food and Environment Research Agency (FERA), but all within tomato-growing glasshouses or packing sheds.

My Evesham moth (Plate 1) apparently represents the first “free flying” example for Britain, at least on the basis of data received by FERA, albeit just a mile or two from a large fruit and vegetable growing area.

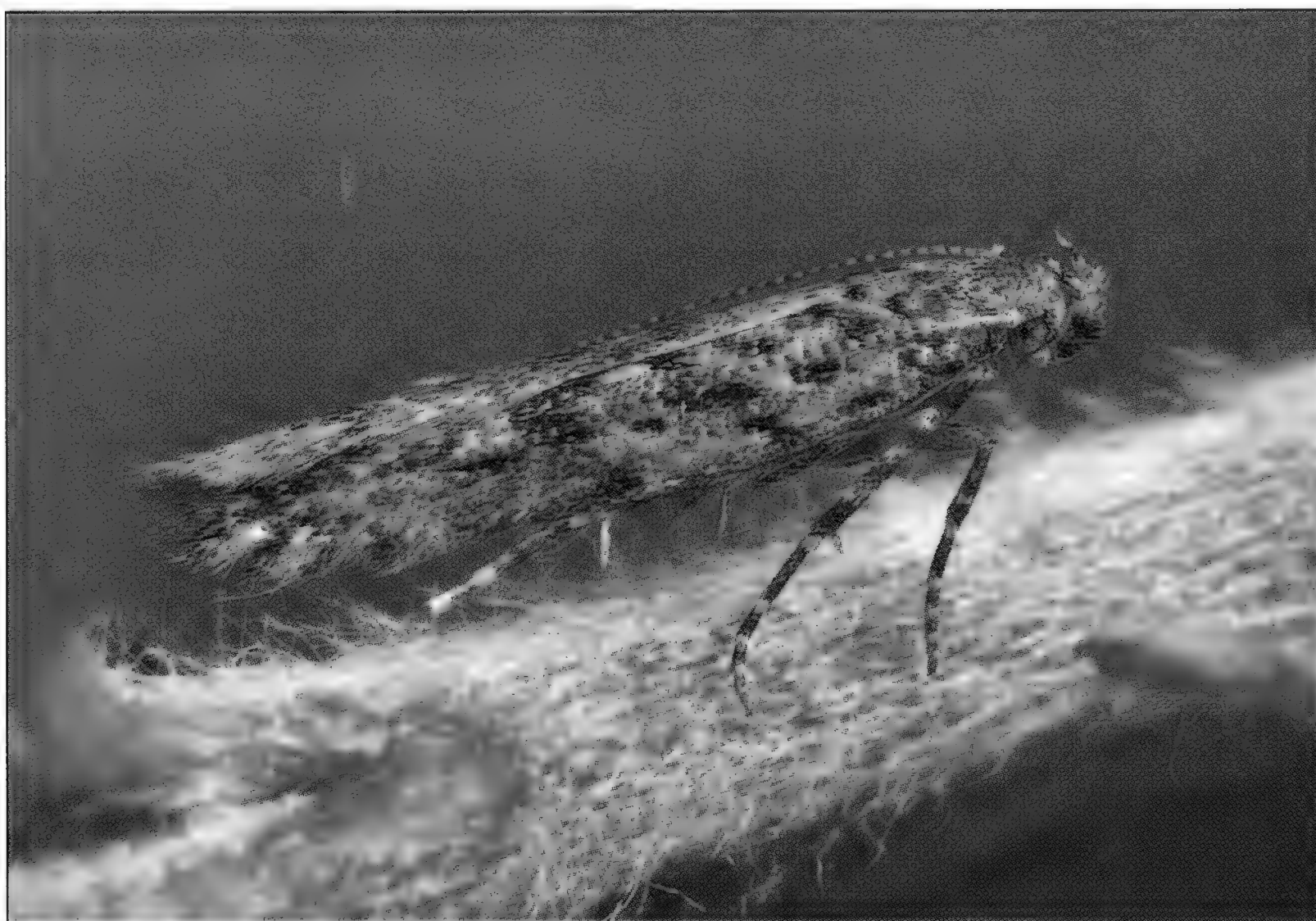


Plate 1. *Tuta absoluta* (Meyr.) Evesham, Worcestershire, July 2010. (Photo: P. Clement)

There seems no reason why this species should not spread into the wider countryside as our climate is apparently suitable and the moth will feed on any solanaceous plants. Only time will tell, but definitely one to watch out for. — PATRICK CLEMENT, 28 Haswell Rd, Halesowen, West Midlands B63 1DA (E-mail: patrick.clement1@btinternet.com).

Unseasonal records of *Apamea monoglypha* (Hufn.) Dark Arches (Lep.: Noctuidae) in Dorset (VC 9)

A presumed single individual of *Apamea monoglypha* Dark Arches was recorded at mv light on the night of 27.xii.2011, 29.xii.2011 and 1.i.2012 at Broadway, Weymouth by Paul Harris; the identification was confirmed from a photograph.

Previous confirmed Dorset records of *A. monoglypha* during the autumn are considered partial second generation, observed mainly at the end of warm summers (P. Davey): Walditch 29.x.2006 (M. Parsons); West Bexington at MV on 5.x.1991, 1.x.2002, 6.x.2003, 6.x.2003 and 18.xi.2006 (R. Eden); 18.x.1997 and 20.x.1997 (P. Sterling and D.Hallett); Lyons Gate 5.x.2005 (T. Box); Puddletown 14.xii.2003 (H. Wood Homer); Iwerne Minster at MV 25.x.1962 (H. Moore), Furzebrook at MV 6.x.1972 (N. Webb); Swanage 11.x.2006 (R. Cox); Gaunts Common at MV 8.x.1991 (P. Davey); St. Ives at MV 3.x.1988 (J. Clarke); Hengistbury Head (Dorset Admin/VC 11) at MV 17.x.1997 (M. Jeffes). — L. J. HILL, 48 East Lodge, East Lulworth, Wareham, Dorset BH20 5QL.

An unseasonal record of *Apamea anceps* (D.& S.) (Lep.: Noctuidae), the Large Nutmeg, in Buckinghamshire (VC 24)

A single example of this noctuid moth was attracted to one of our outside lights on New Years Eve, 31 December 2011. The normal flight period of this single-brooded species in Britain is from early June to mid-July (Skinner, 2009. *Colour identification guide to Moths of the British Isles. Second edition.* Apollo). At a more local level, it flies from the second week of May to the end of July in Buckinghamshire, with a peak in the second week of June and one unseasonal record in November 1972 (Martin Albertini, personal communication). In next-door Hertfordshire, the flight chart given in Plant (2008. *The Moths of Hertfordshire.* HNHS) shows a more or less identical situation (though without any November reports).

Thanks to Peter Hall for confirming the identification of my moth. There are rumours on the moth grapevine of another adult of this same species, also in December 2011, in Berkshire, but I have no details of this. — NIGEL PARTRIDGE, Beechurst, Woodway, Loosley Row, Buckinghamshire HP27 0NS.

December Large Nutmeg *Apamea anceps* (D.& S.) (Lep.: Noctuidae) in Berkshire

A 40-watt actinic trap left to run overnight by Mike A. Wilkins at a garden in The Causeway, Berkshire (VC 22: O. S. grid reference SU 468918) on 27 December 2011 produced a surprising result in the form of a single example of *Apamea anceps*; this was the only moth in the trap. An image of the moth was posted on the Berkshire Moth Group e-group, where its identity was confirmed by Andy Banthorpe and myself.

There are 1,141 previous records of Large Nutmeg on the Berkshire moth database, all between 7 May and 18 August, with the vast majority recorded between late May and mid-July. It is perhaps significant that of the 13 times that Large Nutmeg has been seen *before* 16 May, twelve were in 2011 (the 13th being on 12 May 2002). Three of the early May records in 2011 were at the same site as the 27 December record detailed above.

A few other species have also produced “out-of-date” adults in 2011. A *Phtheochroa rugosana* (Hb.) (Tortricidae) was caught on 9 Nov 2011 at Reading by Jan Haseler; the previous latest date for the county was 10 August and May to July is the norm. A Garden Carpet *Xanthorhoe fluctuata* (L.) came to Les Finch’s trap at Maidenhead on 8 November 2011 (previous latest for the county was 23 October) and a Vine’s Rustic *Hoplodrina ambigua* (D.& S.) at the same site on 26 Nov 2011 (typically no later than September, previous county latest was 6 November). — MARTIN HARVEY, Evermor, Bridge Street, Great Kimble, Aylesbury, Buckinghamshire HP17 9TN (E-mail: kitenetter@googlemail.com).

Unseasonal records of *Apamea anceps* (D.&S.) Large Nutmeg (Lep.: Noctuidae) in Bedfordshire (VC30)

The occurrence is noted of the noctuid moth *Apamea anceps* in Bedfordshire, VC30, in October 2011 followed by no less than nine records in November 2011. The dates and sites are listed here: 30 October at Lower Stondon (A. & M. Banthorpe); 3 & 30 November at Biggleswade (L. Burgess); 9, 12, 18 & 22 November at Upper Caldecote (M. Burgess) and 9 November at Eaton Ford (A. A. Lawrence). There is one other October record for VC30 at Lower Stondon on 18 October 2006 (A. & M. Banthorpe) and two older September records from 1987 and 1993. The usual flight season of this species in the county is from Standard Week 21 (late May) to Standard Week 31 (first week of August) with a few other records in August. It is interesting to note that compared to 2010 the flight period seems to have been nearly three weeks earlier than normal in both its beginning and end with no less than 38 records so far noted in the first two weeks of May (there are only four records from previous years in this date period). It is likely that this earlier emergence due to warm weather, combined with the warm autumn, triggered the emergence of a small second generation of adults. — ANDY AND MELISSA BANTHORPE, 32 Long Close, Lower Stondon, Bedfordshire, SG16 6JS.

Is it early or is it late? A November *Apamea anceps* (D.& S.) (Lep.: Noctuidae) in Hertfordshire

Not to be outdone by our nearby neighbours of Bedfordshire, Buckinghamshire and Berkshire, Hertfordshire too can boast a winter example of *Apamea anceps* — one came to the mv trap run in the adjoining gardens owned by James Fish and Julian Reeves in Bishops Stortford, on the night of 2 November 2011. I am grateful to Jim and Julian for allowing me to mention this here.

The normal flight period of *Apamea anceps* in Hertfordshire, based on 1638 records from 1884 to 2010, inclusive, has been from 16 May to 9 July and so a November record is clearly exceptional. During 2011, however, this “normal” generation was evidently extended with earlier than usual records at Royston, in the north of the county, on 9 and 10 May (John Chainey & Jenny Spence) and later than usual examples in the far west at Aldbury, on 8 August (Gavin Boyd). All of these suggest an extension of the expected single generation of adults; the November record surely cannot be so linked and it remains unresolved whether or not this represents a partial second generation or a confusion caused by the weather triggering responses more normally associated with spring. Given the fact that Blackbirds *Turdus merula* were nest-building in my garden in December, where spring-flowering plants are currently defying the occasional cold spell by thrusting their shoots up above the soil, I suggest the latter is more likely? — COLIN W. PLANT, 14 West Road, Bishops Stortford, Hertfordshire CM23 3QP (E-mail: cpauk1@ntlworld.com).

Unseasonal moths in Northumberland in 2011

In view of the several “southern” records of moths appearing, as adults, exceptionally late during 2011, it seems worth placing on record that the north-east of England was equally affected by whatever unknown factor is inducing these changes. In evidence of this, I trapped a worn Garden Carpet *Xanthorhoe fluctuata* (L.) at light in Tynemouth, Northumberland (VC67) on New Year’s eve, 31 December 2011. All of my remaining records from here for this species fall in the period from April to October.

Other latecomers here include a Brimstone Moth *Opisthagraptis luteolata* (L.) on 13 October 2011 (records usually fall from April to September), a Heart and Dart *Agrotis exclamationis* (L.) on 25 October (typically here from April to August) and two Shuttle-shaped Dart *Agrotis puta* (Hb.) on 4 November (normally fly here April to October).

Two further species might better be described as “early” rather than “late”; a single Hebrew Character *Orthosia gothica* (L.) came to the mv light here on 2 December (normally they never arrive before February) and at the start of the year I had a V-pug *Chloroclystis v-ata* (Haw.) on 6 April (normal flight period July and August). — TOM TAMS, 191 Links Road, Tynemouth, Northumberland (E-mail: tomsphotos@hotmail.co.uk).

Two unseasonal records of *Eupithecia dodoneata* Guenée (Lep.: Geometridae) Oak-tree Pug in Buckinghamshire (VC24)

As of the end of year 2010, of the 298 records of Oak-tree Pug (*Eupithecia dodoneata*) residing in the main Bucks Moth Database which have full dates and are recorded as adults, 63 (21%) were recorded in April, 194 (65%) are records for May, 32 (11%) for June and just 9 (3%) for July. The last date is 23 July (2004)

followed by 18 July (2001). The year 2011 has been a year for plenty of unusual timings, especially following the very warm and dry spring. Recent articles in this journal reflect this with several spring species being recorded in autumn.

On 2 August 2011, at Rammamere Heath (by David Wilton) and 12 August 2011 in my garden here in Ballinger Common, both in Buckinghamshire, individual examples of Oak-tree Pug were encountered. The latter represents the latest record by around three weeks. The moths in question were dissected and identified as a male from Rammamere Heath and a female from Ballinger Common. To quell any lingering doubts about the authenticity of these records, the female's genitalia were photographed and passed over to other experts for absolute confirmation. Thanks to Martin Townsend, Martin Corley, Colin Plant, John Langmaid and Brian Goodey for agreeing with the identification. I am unaware of other late records for this species. Are they really early or really late? Do these examples represent a partial second brood for this normally univoltine moth? Could such a second generation of adults become the norm as seems to be the case with *Eupithecia intricata* in neighbouring Hertfordshire (see Plant, 2008. *The Moths of Hertfordshire*, page 344-245). Take your pick. — PETER R. HALL, Melanthia, Chiltern Road, Ballinger Common, Bucks HP16 9LH.

An unseasonal record of *Orthosia gothica* (L.) Hebrew Character (Lep.: Noctuidae) in Bedfordshire (VC30)

On 21 December 2011 I decided to run my garden MV moth trap for a last time this year. The resulting catch was a complete surprise comprising just a single specimen of a Hebrew Character *Orthosia gothica* (L.). The identity of this specimen was confirmed by county Macro-moth Recorders Andy and Melissa Banthorpe. This record from Clifton, Bedfordshire (VC30, TL 162390) is the earliest/latest recorded for this species in the county and follows on from records from Dorset (VC9) in October reported by Les Hill (*Ent. Rec.* **123**:283). There are two January records for this species in the VC30 dataset: in the Shuttleworth Rothamsted Trap on 7 January 1975 and the Cockayne Hatley Rothamsted Trap on 27 January 1993 (A. & M. Banthorpe pers. comm.). — ALAN R. OUTEN, 14 Fairfax Close, Clifton, Shefford, Bedfordshire SG17 5RH.

Unseasonal Brown Rustic *Rusina ferruginea* Esper (Lep.: Noctuidae) in Middlesex

In the autumn of 2011, I purchased a collapsible 15-watt actinic "Heath" type moth trap to allow me to extend the range of my trapping. On 22 September 2011, I met with Leslie Williams of the London Borough of Brent Parks Service to borrow their portable battery unit and test the trap. We chose Mason's Field, on the edge of Fryent Country Park (Middlesex, VC 21), a former sports ground for which Brent Council were successful in obtaining Heritage Lottery funding for habitat restoration – to improve access and join it up with the adjacent landscape

of meadows and hedgerows. As it was a test event and a little chilly we only trapped for 30 minutes, from 8.00pm. We trapped one Square-spot Rustic *Xestia xanthographa*, two *Tachystola acroxantha* (new for me and the Fryent Country Park list) and a noctuid I initially identified as a Deep-brown Dart *Aporophyla lutulenta* due to the season and presence of a few small whitish marks on each leading edge. On checking with Colin Plant, this moth was revealed to be a Brown Rustic *Rusina ferruginea*, which usually flies from late May to mid-July. A record from 22 September was most unexpected, but does seem to fit a pattern of other unseasonal moths in the autumn of 2011. — DEBBIE PLEDGE, 29 Saltcroft Close, Wembley, Middlesex, HA9 9JJ.

Early appearance of *Xylocampa areola* (Esper) and *Orthosia gothica* (L.) (Lep.: Noctuidae) in northern Spain

Further to the recent reports of spring moth species flying in late autumn/early winter (e.g., Pledge, 2011, *Ent. Rec.*, **123**: 282-283; Hill, 2011, *Ent. Rec.* **123**: 283), it is of interest that this phenomenon may not be confined to Britain. From 24 October until 30 November 2011, SJP caught moths at two to three day intervals (18 night trapping sessions) with a 125-watt MV trap in the Picos de Europa, northern Spain (Linares, Peñarrubia, Cantabria) at an altitude of 420m. During these sessions one fresh-looking *Orthosia gothica* was caught on 19 November and fresh-looking *Xylocampa areola* were recorded on 17, 21 and 26 November. The three *X. areola* were different individuals and not recaptures of the same moth.

Ronkay, Yela & Hreblay (2001, *Noctuidae Europaeae 5: Hadeninae II*: 36) state under the heading of *Orthosia gothica* that “the first adults appear in early March, the flight period usually extending to mid-May”. TF has been undertaking regular winter light trapping from 1993 to 2011 at four nearby locations in Cantabria (Aliezo, Barrio, Cabañes and Pesaguero) and records that the earliest date for an adult *gothica* was 6 February (in 2002). Thus, the November record does appear to be exceptional for this species in the Picos de Europa and is possibly linked to an unusually warm October.

Ronkay et al (*op. cit.*: 290), state that *X. areola* is also univoltine, with the wintering stage most often being the pupa and adults on the wing in early spring. However, they also state that in southern parts [of Europe?] they may appear in late autumn or even during winter. Scrutiny of TF's past data from the Picos de Europa show that *X. areola* was caught in 20 winter trapping sessions, from November-January inclusive, between 1993 and 2011, with the earliest appearance on 5 November 1993.

Thus, the early winter emergence of this latter species appears to be fairly typical, at least as far as recent years are concerned, in spite of the fact that Cantabria lies in the geographical north of Spain and can scarcely be classed as falling within “southern parts”. However, the climate of the area is a complex

mixture of Atlantic, Alpine and Mediterranean influences (Bunce, Bell & Farino, 1998. The Environmentally Sensitive Area legislation in the United Kingdom and its potential application to the Picos de Europa mountains in northwest Spain. *Environmental Conservation* **25**, 219-227), and the latter element may account for the winter flight of *areola*. — TERESA FARINO, Apartado de Correos 59, 39570 Potes, Cantabria, Spain (E-mail: teresa@iberianwildlife.com) and STEVE J. PETTY, Craigielea, Kames, Tighnabruaich, Argyll PA21 2AE (E-mail: stevepetty@btinternet.com)

Early records of *Orthosia gothica* (L.) Hebrew Character (Lep.:Noctuidae) and other “Spring” moths in Yorkshire

In addition to many other counties, Yorkshire had its share of unseasonal moths at the end of 2011. Appearances of *Orthosia* species before the end of the year are not unprecedented in the county (e.g., Talbot, 2001, *Ent. Rec.*, **113**: 17) but are extremely rare, with records of just one Hebrew Character *Orthosia gothica* in 2009 and two Common Quakers *Orthosia cerasi* in 2000 and 2004. To put this into perspective the county database currently holds over 20,000 records of *O. gothica* and 10,000 of *O. cerasi*. On the very early date of 13 October 2011 I was surprised to find a fresh *O. gothica* in my garden trap at Hutton Conyers (VC 65). This was followed by more specimens of the same species on 3 November, 22 December and 23 December. Another was trapped nearby at Sharow (VC 64) on 31 December.

These were not the only “Spring” moths to be reported in the county at the end of 2011. Common Quakers *Orthosia cerasi* were trapped by John Walshe in Leeds on 27 and 31 December and a Spring Usher *Agriopis leucophaearia* was trapped by Stephen Sowden on Hatfield Moors on 31 December, the latter being the earliest record ever in the county. Several Pale Brindled Beauties *Phigalia pilosaria* were also reported before the year end, but this has become an annual event since 2000 with the exception of the last two very cold winters when none were reported.

Most people trapping moths tend to put away their traps at the onset of winter and only a few “die hard” enthusiasts continue to trap through the coldest months, so there were probably many more early species on the wing than these records suggest. — CHARLES FLETCHER, The Forge, Hutton Conyers, North Yorkshire (E-mail: chfletcher@btinternet.com).

First record of the Large Thorn *Ennomos autumnaria* Werneb. (Lep.: Geometridae) in Leicestershire

During the period of 10-13 September 2010 a single specimen of *Ennomos autumnaria* was caught in the Rothamsted Insect Survey light-trap at Loddington (site 560: O. S. grid reference SK 792024). Whilst not new to VC 55, which incorporates Rutland, this record was the first for Leicestershire. Only ten miles to the east of Loddington is Barrowden (Rutland), where the same garden has provided

the four VC 55 *E. autumnaria* records – on 25 September 2000, 3 September 2005 and 8 and 19 September 2008. This Nationally Scarce species is predominantly restricted to south-east England, where it spreads from strongholds in Bedfordshire and Cambridgeshire Hertfordshire, Norfolk, Suffolk, Essex, Kent, Sussex and Hampshire (Hill, L., Randle, Z., Fox, R. & Parsons. 2010. *Provisional Atlas of the UK's Larger Moths*. Butterfly Conservation), but perhaps this record indicates a range expansion, a process which has been evident since the 19th century. The species inhabits broad-leaved woodland, scrub and gardens where the larvae feed upon a wide range of trees and shrubs (Waring, P., Townsend, M. & Lewington, R. 2009. *Field Guide to the Moths of Great Britain & Ireland*. British Wildlife Publishing), so it is therefore surprising that it is not more widely distributed.

Many thanks to John Szczur at Loddington for bringing the importance of this record to my attention and for operating the light-trap so well and to Adrian Russell, the VC55 recorder, for confirming its status and for extra information. — PHILIP J. L. GOULD, Co-ordinator of the Rothamsted Insect Survey Light-trap Network, Plant & Invertebrate Ecology Department, Rothamsted Research, Harpenden, Hertfordshire AL5 2JQ (E-mail: phil.gould@rothamsted.ac.uk).

Thyme Pug *Eupithecia distinctaria* (H.-S.) (Lep.: Geometridae). Why is it so local?

Thyme Pug *Eupithecia distinctaria* has a wide range in Britain, from Dorset on the south coast of England to the Outer Hebrides. However, it is apparently absent from the whole of eastern Britain. Recent records are mainly western and coastal (Hill, L., Randle, Z., Fox, R. & Parsons, M., 2010. *Provisional Atlas of the UK's Larger Moths*. Butterfly Conservation, Dorset). They are also sparse. In total, the atlas dot map for this species shows records in 85 10km squares, but just 23 (27%) are post-2000 sightings, despite the increase in recording effort generated by the National Macro-moth Recording Scheme. Many of the 62 pre-2000 dots are well inland and there must be some doubt whether all these records are correct, since pugs were so often misidentified before the wealth of modern field guides.

As its English name implies, Thyme Pug is dependent upon Wild Thyme *Thymus polytrichus* in Britain. Its caterpillar feeds only upon the flowers. Riley & Prior (2003. *British and Irish Pug Moths*. Harley Books, Colchester) list no other foodplant in the wild, nor do any other authors. Yet there is an anomaly here. All British authorities agree that Thyme Pug is on the wing in June and July, with the caterpillar found in August and September. Porter (1997. *The Colour Identification Guide to Caterpillars of the British Isles*. Viking, London) is slightly more specific, giving the larval stage as late July to mid September. Yet floras such as Garrard & Streeter (1983. *The Wild Flowers of the British Isles*. Macmillan, London) give the flowering period of Wild Thyme as from May to July. That is rather awkward for a larva that feeds in August and September.

My first acquaintance with Thyme Pug came on Ardnamurchan (West Inverness-shire) during 7-9 July 1997. Adults were readily disturbed in sunny

weather from rocks or put up from the short flowery turf where Wild Thyme flourished. None was in a suitable condition for photography, so I took a female for eggs in the hope of breeding better specimens.

The resulting caterpillars did well on the flowers of a taller, non-native species of *Thymus* from the herb garden. By the first week in August they were in their final instar, approaching full size. Then the thyme stopped flowering. At first I was not particularly concerned. Surely the caterpillars would complete their growth on the young seedpods or some other part of the plant? No, they did not. I realised too late that only flowers would do.

This failure rankled, making me eager to try again. It was a long wait for the next opportunity. Then on 20 July 2011 John Kemp trapped four Thyme Pugs on Eriskay in the Outer Hebrides. He very kindly obtained eggs and posted them to me. They hatched after about 10 days and the tiny yellow larvae duly began feeding on my garden *Thymus*. This was already coming to the end of its flowering period, presaging the same problem as before. Another source of foodplant was urgently required. Wild Thyme grows abundantly in various places on the Banffshire coast, but a visit on 3 August found that this too was virtually over. Only the occasional cushion still bore a couple of flowers. Things were looking serious! However, Allan (1949. *Larval Foodplants*. Watkins & Doncaster, London) gives flowers of Marjoram *Origanum vulgare* as an acceptable alternative in captivity. I mixed this with what was left of the garden thyme and after some initial reluctance it was accepted.

Both thyme and Marjoram flowers were eaten in a similar and highly selective way. A caterpillar inserted its head into the throat of a flower and ate the pistil, stamens and lowest part of the corolla tube. The upper part of the corolla was not eaten, but fell to the ground when detached. Soon the bottom of the container became carpeted with pink petals, like fallen blossom under a cherry tree. Clearly the caterpillars were selecting only the most nutritious parts of the flower. They grew quickly on this diet, becoming fully fed in three to four weeks and descending to the floor of the container. I removed each one in turn and provided dry sphagnum moss for pupation. Only then did I realise exactly how many there were, so well-camouflaged had they been amongst the thyme and Marjoram flowers. Indeed, when renewing the supply I never discarded the old foodplant until frass stopped appearing beneath it. Invariably I had missed a good few larvae, even in the final instar, despite the most meticulous search. My earliest count had detected only six caterpillars, but the final tally was 35. That's quite embarrassing, really!

Does this dependence on the flowers of thyme, yet a larval stage when flowering is mainly over, explain why Thyme Pug is so very local in Britain? Can it only survive in areas where thyme carries on flowering later than usual, through August and into September? There is plenty of apparently ideal habitat on the Banffshire coast, for instance, yet thyme flowers were virtually finished before the

end of July in 2011. No wonder the moth is absent. But how about Eriskay, the source of my captive stock? On 22 August John Kemp went back to the trapping site to check. He found just two small patches of thyme with a few flowers left, the rest being over. It seems likely that my captive larvae would have perished for lack of food had the eggs been laid in the wild, as the larval stage would almost certainly have lasted longer than in captivity.

Tom Prescott (pers. comm.) has found that in East and West Inverness-shire some Wild Thyme plants along the edges of paths and tracks will carry on flowering at least until the end of August if they are subject to trampling or grazing pressure, whereas undamaged plants in the same area have long finished. Perhaps it is these conditions that the moth requires. Even so, it is rather unusual for a species to have a life history that is out of synchronisation with its sole foodplant. Moreover, in Thyme Pug's case it seems unnecessary. Why does the adult fly so late? Why not emerge in May (as do so many of the *Eupithecia* species that overwinter as a pupa) when its foodplant is just starting to flower? That would give its caterpillar ample opportunity to feed up. There must be strong factors working against this, otherwise natural selection would have brought it about. Maybe as a consequence, this species is much more scarce and local in Britain than we might expect from the distribution of its foodplant. Perhaps the answers to these puzzles lie in its European range. Thyme Pug has a classic Mediterranean distribution (Mironov, V., 2003. *The Geometrid Moths of Europe*, Vol. 4. Apollo Books, Stenstrup). Thus in the British Isles it is very much at its north-western limit. As with other such species, in order to survive here at all it may have particularly exacting habitat requirements. Elsewhere in Europe it has a far wider range of foodplants including Perforate St John's-wort *Hypericum perforatum*, Slender Bedstraw *Galium pumilum* and non-British *Odontites* and *Cytisus* species (Leraut, P., 2009. *Moths of Europe* vol. 2. N. A. P. Editions). Presumably the timing of its larval stage is less of an issue there.

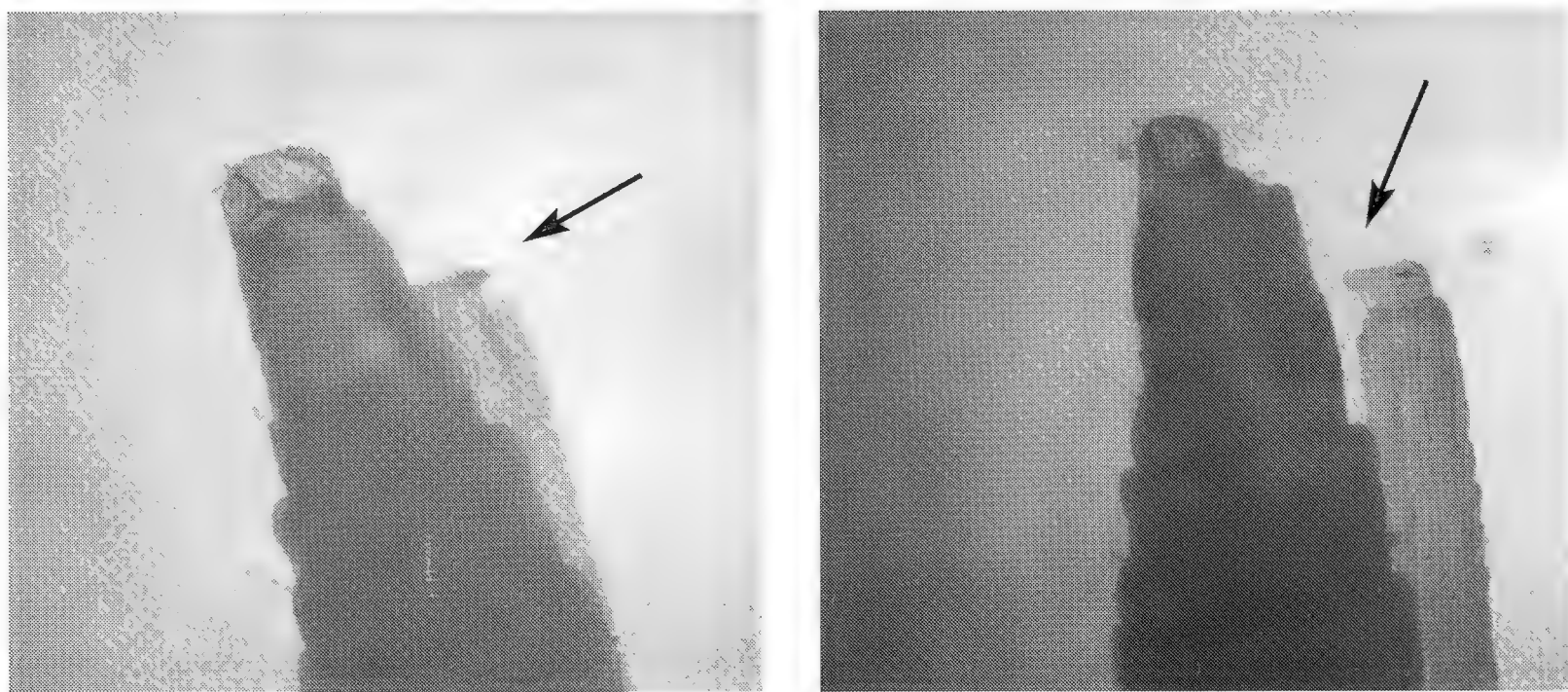
I am grateful to John Kemp for sending me the eggs and for making further observations on Eriskay, also to Tom Prescott for information on late-flowering thyme. Some useful comment was received via the ScottishMoths Forum.— ROY LEVERTON, Whitewells, Ordiquhill, Cornhill, Banffshire AB45 2HS.

Phyllonorycter leucographella* (Zeller, 1850) (Lep.: Gracillariidae) and *Prunus

On 22 September 2011, I found and photographed some examples of upperside blotch mines on some low branches of a sucker from a cherry tree (*Prunus* sp.) in suburban Cheltenham. The mines were centred on either the midrib or side veins and were for the most part elongated and narrow ovoids in shape, a typical example measuring 26 mm on the long axis over the midrib and 5 mm maximum width. The mines were a silver white in colour and to all intents looked like those of *Phyllonorycter leucographella* (Zell.). However, leaf miners on rosaceous plants are difficult to identify from mine morphology alone and it seemed sensible to try

eliminate two other strong contenders – *Phyllonorycter cerasicolella* (H.-S.), which feeds on cherry, albeit in mines on the underside of the leaves and *Phyllonorycter corylifoliella* (Hb.), which feeds on a range of rosaceous shrubs with generally rounded upperside blotch mines that ultimately are silver white in colour with brown speckling.

Opening one of the mines produced a long-dead caterpillar, 1.4 mm in length; a second example contained only granules of black frass, but a third mine contained a very lively pupa. The cremaster had two pairs of spines, the outermost pair being broad at the base, triangular in shape and each narrowing to a curved point, with that point directed outwards; the inner pair were more elongate and the points were



Figures 1 and 2. Apical spines on the developing legs of the Cheltenham pupa.

directed inwards. The impression created was of two pairs of strongly diverging spines. This description accords with illustrations of the *leucographella* cremaster in, for example, Patocka J. and Turcani M. (2005. *Lepidoptera Pupae*. Apollo). However, Triberti (2007. The *Phyllonorycter* species from Palearctic Region feeding on Rosaceae (Lepidoptera, Gracillariidae). *Bollettino del Museo Civico di Storia Naturale di Verona* **31**, *Botanica Zoologia*: 147-221) warns against a reliance on cremaster features to differentiate between rosaceous *Phyllonorycter* species 'as shape, size and position of the cremaster pairs of spines are very similar and variable'. Fortunately, the pupa of *leucographella* does have another characteristic with which an identification can be secured, in the form of strongly hook-shaped apical spines on the hind legs, illustrated by Triberti on page 193. These were clearly visibly on the Cheltenham example (Figs. 1 and 2). The cremaster and, above all, the hind-leg spines indicate that the mine examined was indeed that of *Phyllonorycter leucographella* and the Cheltenham example constitutes the first confirmed instance of the species feeding on *Prunus* in Britain.

I am grateful to Rob Edmunds for his comments on photographs of the mines and pupa made available on the Yahoo ukleafminers discussion group web site.
— ROBERT HOMAN, The Apiary, Swindon Lane, Cheltenham, Gloucestershire GL50 4PD.

***Stephanitis takeyai* Drake & Maa (Hem., Tingidae) at Box Hill, Surrey**

On 19 August 2009 I paid a visit to Box Hill, Surrey, to seek two of the species of Hemiptera for which this site is noted: *Gonocerus acuteangulatus* (Goeze) (Coreidae) and *Anthocoris butleri* Le Quesne (Anthocoridae), both associated with box (*Buxus sempervirens* Linnaeus, Buxaceae). Leaving my vehicle in the Zig Zag car park (grid reference TQ176519), I ascended the adjacent hill-side to the box hedge, and started to beat. Both target species were found very easily, and in the course of my search I was delighted to find also a black and white patterned lacebug (Tingidae), somewhat resembling a small bird dropping, which was clearly of a species unfamiliar to me. At home that evening, using my copy of Southwood and Leston (1959, *Land and Water Bugs of the British Isles*, Warne, London), the insect keyed out as *Stephanitis rhododendri* Horvath (Tingidae), a species I had been hoping to find for some years, although hitherto without success. The absence of *Rhododendron* spp. (Ericaceae) at the site, the host plants of this species, was a little puzzling, but I did not question the matter further. Four days later, I returned to Box Hill to find more examples of the insect, but in spite of a prolonged search I encountered only another singleton, this time by sweeping low vegetation around the edge of the car park.

There the matter remained, until a few days ago when I was reading some back issues of the *British Journal of Entomology and Natural History*. I came across the paper of Halstead and Malumphy (2003, 16: 3-6) describing the introduction to Britain of *Stephanitis takeyai* Drake & Maa with the ornamental shrub, *Pieris japonica* (Thunb.) D. Don (Ericaceae), a pest of this plant. The bug was reported from two gardens, elsewhere in Surrey, and the paper included some excellent images of the insect. I immediately remembered my specimens from two years before, and a re-examination of these revealed that I had made a mistake: the insects were *S. takeyai*, not *S. rhododendri*. The plant association of my specimens is still wrong, but I presume that the insects were in the process of dispersing from a nearby site.

I reported my find to the DEFRA Plant Health and Seeds Inspectorate, who informed me that they are no longer trying to eradicate this pest from Britain, and that the insect is now considered established. By 2006, the bug had been reported from approximately 30 plant centres, and attempts to remove the insect were thwarted by prompt re-colonisation from elsewhere. No doubt, sightings of the bug will continue in parks and gardens, but it will be interesting to see how the insect establishes itself in the countryside, given that the bug can also infest *Rhododendron* spp., which grow wild in many woodland areas.

In addition to the above paper, more images of this bug may be found on the British Bugs website (<http://www.britishbugs.org.uk>) or by searching Flickr (<http://www.flickr.com>). — ROB RYAN, 38 St John Street, Oxford, OX1 2LH (E-mail: notnowcato@ymail.com).

New records of Triple-spotted Pug *Eupithecia trisignaria* H.-S. (Lep.: Geometridae) in Bedfordshire, VC 30

On the night of 10-11 July 2009 the Rothamsted Insect Survey light-trap at Cockayne Hatley (site 336: O.S. grid reference TL 253494) caught a single specimen of *Eupithecia trisignaria*. This was only the second record for VC30, following a single male found at Maulden Wood by Colin Plant on 6 August 2003. Further records then followed in 2010 when A. M. and M. G. Banthorpe found a pair in Maulden Wood on 21 July; and then a male and three females in Chicksands Wood on 8 August.

E. trisignaria inhabits roadside verges, woodland rides, marshy fields and fenland, where the larvae feed upon the ripening seeds of Wild Angelica, *Angelica sylvestris* and Hogweed *Heracleum sphondylium* (Skinner, B. 2009. *Colour Identification Guide to Moths of the British Isles*. Apollo). It is widespread but local in England and Wales and very local in Scotland; it is also locally common in the north of Ireland (Hill, L., Randle, Z., Fox, R. & Parsons, M., 2010. *Provisional Atlas of the UK's Larger Moths*. Butterfly Conservation).

It seems likely that *E. trisignaria* has been over-looked in Bedfordshire and that populations will be found in other well-managed woodlands where there are good rides containing the foodplant (A. Banthorpe, pers. com.).

My thanks to Andy and Melissa Banthorpe, the Bedfordshire recorders, for letting me know about the importance of the Rothamsted record and for further information; and to Ian Woiwod for his continued operation of the light-trap and identification of the catches.— PHILIP J. L. GOULD, Co-ordinator of the Rothamsted Insect Survey Light-trap Network, Plant & Invertebrate Ecology Department, Rothamsted Research, Harpenden, Hertfordshire AL5 2JQ. (E-mail: phil.gould@rothamsted.ac.uk).

Unusual refreshment for an Old Lady . . . and others

Roger Hayward (*Ent. Rec.*, 123: 221), noted an Old Lady *Mormo maura* (L) (Lep.: Noctuidae) drinking from his pond. I can confirm that other species of moth are quite partial to pond water – it is a trick I discovered whilst manning the Lincolnshire Naturalists' Union stand at the annual Lincolnshire Show. A number of moths (commonly the hawk-moths – Sphingidae) are captured the night before and kept cool for display during the show. After a few hours of being very cooperative “on display” (much to the amazement of the general public) they start to get slightly agitated. I have found that a few drops of pond water from the “pond creatures” tank is enough to settle them down. — CHARLIE BARNES, East Cottage, Nocton Rise, Lincoln, Lincolnshire LN4 2AF.

The Rothamsted Insect Survey light-trap on Jersey – specimens of note from 2009

There have been Rothamsted Insect Survey light-traps operating on Jersey, in the Channel Islands, since 31 December 1969. The first trap at Howard Davis Farm, Trinity ran until 1993, but then had to be moved about 100m to its current position (Jersey II: site 547). This long period of consistent trapping has provided one of the Insect Survey's largest and most valuable long-term data sets. Not surprisingly, given its proximity to France, every year the trap turns up moths from Europe that would be considered uncommon on the British mainland. However, despite the length of time it has been running, the trap still catches species that are of particular interest.

On the night of 11/12 May a single specimen of the Dwarf Pug *Eupithecia tantillaria* Boisd. (Geometridae) was caught. This is the first record of this species for Jersey. The only other record for the Channel Islands was caught in an MV trap at La Vallée on Alderney on 26 April 2007. The larvae of *E. tantillaria* are polyphagous on several species of native and introduced conifers, particularly favouring Norway Spruce *Picea abies*, Sitka Spruce *Picea sitchensis* and Douglas Fir *Pseudotsuga menziesii*. The species can be found not just in plantations but also where conifers occur in parks and gardens (Waring, P., Townsend, M. & Lewington, R. 2009. *Field Guide to the Moths of Great Britain & Ireland*. British Wildlife Publishing). It is, therefore, surprising that it has not been recorded more frequently on the Channel Islands, as they fall well within the geographical range shown by Mirinov (2003. *The Geometrid Moths of Europe* 4. Apollo Books). So, despite the fact that individuals from France could be recorded, perhaps *E. tantillaria* is also a very scarce and under-recorded resident on the Islands.

Over the weekend of 12-15 June a Brindled White-spot *Parectropis similaria* Hufn. (Geometridae) came to the trap. This is only the second record for Jersey, after the first was caught at Grouville on 19 July 2006. Elsewhere on the Islands, two were recorded on Alderney in David Wedd's MV trap at La Vallée on the nights of 3 and 4 June 2006. The summer of 2006 was outstanding for migrants and both of these nights were very hot with slight easterly winds (D. Wedd, pers. com.), so it is very likely that these individuals, and Jersey's July record, were migrants. Similarly, given that there have been no further records, it would seem probable that this second specimen for Jersey also originated elsewhere. The species has not been recorded on Guernsey. *P. similaria* is locally widespread in southern England, with populations extending into Devon, north-east to Norfolk and north-west towards Denbighshire (Skinner, B. 2009. *Colour Identification Guide to Moths of the British Isles*. Apollo Books). Hill, L., Randle, Z., Fox, R. & Parsons, M. (2010. *Provisional Atlas of the UK's Larger Moths*. Butterfly Conservation) show the distribution of *P. similaria* as fairly widespread but strictly confined to the south-east of a line drawn between the Severn Estuary and The Wash, perhaps with a tendency to follow calcareous soils, and extending marginally north-west from here into Herefordshire, Worcestershire and Warwickshire. It is, however, perhaps surprising that *P. similaria* is not more

common, as the larvae feed on a wide range of shrubs and trees including oaks *Quercus* spp., hawthorns *Crataegus* spp. and birches *Betula* spp. that are not so limited.

Another second record for Jersey was that of the *Uresiphita polygonalis* D. & S. (Crambidae) specimen caught during the period of 26-31 August. The Channel Islands' first was found by T. Peet at Le Chêne, Guernsey on 24 August 1983. This was followed almost exactly 21 years later by the first for Jersey, at Grouville Common on 21 August 2004. David Wedd then recorded the first for Alderney on 17 October 2005 at La Vallée with a second found at the same site on 28 July 2006. Guernsey's second and third records also occurred in that excellent migration year, with one at St. Peter and the other at St. Martin on 1 and 23 September respectively. Since the Rothamsted record, one more has been found on Guernsey, by Rich Austin at Dell Nursery on 12 October 2010.

This is quite a large 'pyralid' (wingspan up to 37mm), yet it still looks rather modest, at least until the exposure of the striking orange hindwings with their dark border. As on the Channel Islands, it is a very scarce migrant to the British mainland, where it has mainly been recorded along the south coast. The larvae feed on various low-growing herbaceous plants and the species is found from central and southern Europe, through the tropics to Australia (Goater, 1986. *British Pyralid Moths*. Harley Books). Given that it has become a little more frequent in recent years, perhaps a warming climate and the ubiquity of foodplants will allow it to become more resident on the Channel Islands and in southern Britain in the future?

Many thanks to Roger Long (Chairman of the Entomology Section, Société Jersiaise) for his assistance in identifying these interesting species and for further information. Thanks also to David Wedd and Rich Austin regarding Alderney and Guernsey records, respectively, and to Alex Vautier for her long-standing hard work operating the light trap.— PHILIP J. L. GOULD, Co-ordinator of the Rothamsted Insect Survey Light-trap Network, Plant & Invertebrate Ecology Department, Rothamsted Research, Harpenden, Hertfordshire AL5 2JQ (E-mail: phil.gould@rothamsted.ac.uk).

The larval stage of *Epagoge grotiana* (Fabricius) (Lep.: Tortricidae)

On 5 May 1996 I reared *Epagoge grotiana* (Fabricius), Tortricidae, from a larva that I had collected on 2 March that year at Havant Thicket, Hampshire in the company of Dr J. R. Langmaid. As a result of chancing upon an article by Bainbrigge Fletcher (1939. *Entomologist's Rec. J. Var.* **51**: 169-171) and consequent consideration of other British literature, it appears that the larval stage may not have been previously observed in the British Isles or if it has, it has not been published.

Bainbrigge Fletcher (*loc. cit.*) states that the early stages seem to be very little known in England. He reviews various publications between 1905 and 1928, including a note by Huggins (1925. *The Entomologist* **58**: 98) who comments that 'The food-plant is at present unknown, but I should think ... that it is certainly oak,

as I took no fewer than nine specimens, including three of the sluggish female, newly emerged on a single oak bush ...'. Bainbrigge Fletcher drew attention to an apparently overlooked paper by Chrétien (1897. *Le Naturaliste* **19**: 258-260). This gives a very full account of the early stages from ovum to pupa that Bainbrigge Fletcher summarises. He states that Chrétien records that the young larva spins a tube of white silk and feeds on the ground on fallen leaves, preferring discoloured and dry ones. He comments that it grows very slowly and that towards the end of October, when still only about half-grown, it ceases to feed and hibernates in a leaf-fold lined with silk. In March it recommences feeding on vegetable rubbish, dead leaves and dead insects, as well as on fresh leaves of low plants.

Bainbrigge Fletcher then provides a translation of Chrétien's larval description, as follows:

'The full-grown larva is about 14 x 2 mm., slightly attenuated at extremities, with well-marked segments: colour a livid brownish-grey, darker dorsally, each segment with two darker transverse dorsal bands, the anterior one broader, paler on sides, beneath and on incisions; warts very indistinct, of the ground-colour, small and (at least the trapezoidals) with a minute blackish dot emitting a pale hair; head rather flattened anteriorly, rounded on vertex, of a bright honey-yellow, suffused yellowish-brown towards epistome, ocelli blackish; a dark rusty-brown wedge-shaped lateral nuchal marking; prothoracic plate anteriorly concolorous with head, posteriorly and laterally blackish-brown; anal plate brown.'

Bradley, Tremewan & Smith (1973. *British Tortricoid Moths*, Cochylidae and Tortricidae: Tortricinae: 135-136) provide the following larval description: 'Head brown with black ocellar and epistomal regions, or entirely black; abdomen green or greenish brown; pinacula small, black; thoracic legs black.' They give the larval foodplants as the leaves of *Quercus*, *Crateagus* and *Rubus* and the larval period as August to May, overwintering in a narrow fold of a leaf. They go on to state, 'On the Continent it is recorded as polyphagous on low plants, including *Petasites alba*, *Rosa* and *Vaccinium*; also on fallen leaves and fruit'. This rather suggests that it had not been recorded in the British Isles on fallen leaves.

This larval description is less detailed than that given by Bainbrigge Fletcher (*loc. cit.*) and the colour and markings of the head and abdomen differ from his translation of Chrétien's account, thus suggesting that Bradley, Tremewan & Smith did not follow this. If their description was based on a publication in the British literature then I have not been able to trace this.

Emmet (1988. *A field guide to the smaller British Lepidoptera*: 161; 1991. In Emmet, A. M. & Heath, J. (Eds) *The Moths and Butterflies of Great Britain and Ireland* **7(2)**: 146-147) cites the same foodplants as Bradley, Tremewan & Smith and in the former states that the larva feeds on the leaves and overwinters in a narrow fold of leaf. In neither account is there any suggestion that the larva feeds on dead leaves, unlike, for example, the entries for *Clepsis consimilana* (Hübner).

The larva that I found was amongst dead oak leaves on the ground. It was within part of an oak leaf that it had formed almost into a cone shape and initially it fed by protruding its head from this 'case'. It later abandoned this case and fed

externally on dead oak leaves before pupating amongst them. Unfortunately I made no larval description. — R. J. HECKFORD, 67 Newnham Road, Plympton, Plymouth, Devon PL7 4AW.

***Tuponia brevirostris* Reuter (Hem.: Miridae) at Lepe, Hampshire**

There are two species of *Tuponia* Reuter resident in Britain, both of which live on Tamarisk (*Tamarix spp.*, Tamaricaceae), an ornamental shrub of parks and gardens, which also grows wild in many places on the coasts of southern England. *T. mixticolor* (A. Costa) (formerly known as *T. carayoni* Wagner) is of a chequered brown appearance, 3.0-3.5mm long, and was first found in Britain in 1979 at two sites 25km apart on the Solent coast: Freshwater, Isle of Wight and Hill Head, Lee-on-Solent, Hampshire (Nau, 1980, *Entomologist's Monthly Magazine*, 116:83-84). *T. brevirostris* Reuter is of a uniform light green colour, a little smaller at 2.5-3.0mm long, and was first recorded at Chelsea Harbour, Middlesex in 2001 (Barclay and Nau, 2003, *Entomologist's Monthly Magazine*, 139:176-177). Immediately following the discovery of *T. brevirostris*, a wide search was made to find further sites, in Cornwall, Hampshire, Kent and Suffolk, but without success. However, *T. mixticolor* was found in all these counties, except Cornwall, and had previously been recorded from Dorset and Devon, showing how far this bug had spread in the previous two decades.

The initial spread of *T. brevirostris* seemed very rapid. In 2003, additional sites were found at Church Norton on the West Sussex coast (Denton, *Het News*, Spring 2004, 3:5) and at Potters Bar, Hertfordshire (Widgery, *Het News*, Autumn 2004, 4:7-8). I can report that the insect has now reached the area in which its congener was originally discovered, at Lepe Country Park on the Solent coast, almost midway between the two 1979 sites. Both species of *Tuponia* were found on 12 July 2010, in very large numbers, on an extensive stand of Tamarisk growing out of a low cliff, a short distance east of the Lepe Beach Cafe (grid reference SZ 459986). The following month, similarly large numbers were also found at West Wittering, West Sussex, on a long stand of Tamarisk acting as a wind-break between the beach and the car park (11th August 2010, SZ 765984). This second site is just 10km west of the 2003 Church Norton location.

Given the initial rapid rate of expansion of *T. brevirostris*, it is probable that this insect has been in Hampshire for some time, and it would be interesting to know the current extent of its westwards and northwards expansion. Nau (1980, *op. cit.*) comments that Tamarisk is rather scarce on the coast west of Southampton, and *T. mixticolor* had not been found in Cornwall in 2001. Are the ranges of these two bugs now concordant, or are there sites supporting one species but not the other?

Some excellent images of these bugs can be found on the British Bugs website (<http://www.britishbugs.org.uk>) or by searching Flickr (<http://www.flickr.com>). — ROB RYAN, 38 St John Street, Oxford, OX1 2LH (E-mail: notnowcato@ymail.com).

ANNOUNCEMENTS

AWARD FOR INVERTEBRATE CONSERVATION

The Editor and his colleagues feel that readers will wish to join them in congratulating Peter Harvey, of the Essex Field Club, on being awarded an Honorary Doctorate from the University of Essex. Whilst Peter may not be widely known in Lepidoptera circles, we feel it pertinent to mention in this journal that the honour has been bestowed, primarily, in recognition of Peter's outstanding contribution to recording invertebrates and the interpretation of their significance, particularly on the many post-industrial and other brownfield sites within the East Thames Corridor, and elsewhere, over the past twenty or so years.

It should be added that Peter's work has been on an exclusively voluntary basis, something that is entirely in keeping with the traditional amateur aspect of entomological research in Britain. We welcome the recognition of the enormous importance of such unpaid input. I hope that it is not seen as any kind of distraction from this honour that I now also mention that it is, sadly, a fact that there are, outside purely commercial ventures which tend to be fairly superficial, very few opportunities for paid field survey of invertebrates in Britain. Almost all invertebrate conservation planning depends exclusively on the work done by amateurs; with that in mind it is of interest to observe the number of short Notes in this issue of the journal reporting the amateur findings of moths outside their normal flight period – results that may be of some interest to professional climate change researchers. It is hard to see any change to this situation given the current financial climate, but we nevertheless hope that any politicians reading this may take note!

IS THE DIAMOND-BACK MOTH MARKED FOR EXTINCTION?

The Department for Environment, Food and Rural Affairs (Defra) is considering plans by British company Oxitec for the 'open release' of a genetically modified strain of the diamond-back moth *Plutella xylostella* (L.) (Yponomeutidae), whose larvae feed on cabbages, broccoli, cauliflowers and similar crops. The strain is already available and has a lethal gene inserted in the male so that when they mate with wild females their offspring die almost immediately, theoretically causing the population to crash. The intention is to increase crop yields. Oxitec is keen to begin 'trials' next year, but faces opposition from groups who say the untested technology could threaten wildlife and human health. Oxitec says all the GM moths carry a lethal gene and would die within a few days of release. This is known as 'biological containment' and Oxitec argues that it is so successful there is no need for any physical barriers to stop the insects flying away. A Defra spokesman said that while its officials and advisers have discussed Oxitec's plan, the department has not reached a view on whether it should go ahead.

NEW LEAFROLLERS (LEP.: TORTRICIDAE) FOR BULGARIA WITH TAXONOMICAL COMMENTS

BOYAN ZLATKOV & OGNJAN SIVILOV

Sofia University "St. Kliment Ohridski", Faculty of Biology, Dept. of Zoology and Anthropology,
8 Dragan Tsankov Blvd., BG-1164 Sofia, Bulgaria
(E-mail: bzlatkov@gmail.com or osivilov@gmail.com)

Abstract

Seventeen species of Tortricidae are reported from Bulgaria for the first time. One of these, *Dichrorampha caucasica* (Danilevsky, 1948), is also new for the Balkan Peninsula. Some taxonomical comments on this species are added. Everted vesicae of two species are described and illustrated for the first time.

Keywords: Faunistics, Endangered habitats, genitalia, Cochyliomorpha, Pammene, Dichrorampha.

Introduction

Active moth collecting has been undertaken by the authors in a variety of localities across Bulgaria, mostly in the south-western part of the country, during last few years. As a result some papers on Tortricidae are already published (Zlatkov, 2008; Zlatkov & Budashkin, 2010). Field work has continued and a number of species new to the country have been discovered. In this paper seventeen species from Bulgaria are reported for the first time. Many of species reported here are "southern" and have Mediterranean or related type of distribution. This is perhaps the reason why most of them are found in south-west Bulgaria.

Some specimens from two national collections were identified, revealing some species have not been reported previously because of misidentification (e.g. *Acleris napaea*). It should be mentioned that a number of Tortricidae specimens in the collection of the Natural History Museum (Sofia) are incorrectly determined because they have not yet been subjected to genitalia dissection. More new species for the Bulgarian fauna may thus remain undetected whilst, of course, some published data may be incorrect.

The Bulgarian Tortricidae fauna is in any event poorly recorded. MV light trapping is a relatively new concept for our country and very few Bulgarian entomologists collect Microlepidoptera using these methods. Some habitats, especially marshes and wetlands, the Black Sea coast line and others have been particularly neglected for many years. Several species that are common in Europe probably remain, still, undetected here.

Material and methods

Vesica eversions were made using the technique described by Zlatkov (2011). Fresh material, collected by the authors, is preserved in the collection of Boyan Zlatkov in the Faculty of Biology, Sofia University (FBBZ). For existing material,

from the Institute of Biodiversity and Ecosystem Research (IBER; formerly Institute of Zoology, IZ) and the National Museum of Natural History in Sofia (NMNH) the collecting method is noted if mentioned in the label. Collection localities are given with their coordinates when they are available.

Results and discussion

Acleris napaea (Meyrick, 1912)

Bulgaria: SW Bulgaria, Kresna Gorge, Stara Kresna railway station, 09-10.v.1977, 1 ♀, leg. A. Slivov (IBER); near the town of Kresna, 05.vi.1930, 1 ♂, leg. K. Tuleshkov (NMNH), incorrectly determined as *A. rufana* ([Denis & Schiffermüller], 1775), det. H. Rebel. **New for Bulgaria.**

Distribution: This Turano-Iranian species is known from Iran, Pakistan, Afghanistan, Uzbekistan, Armenia, Turkmenistan, Southern Russia (Razowski, 2002).

Phtheochroa annae Huemer, 1990

Bulgaria: NW Bulgaria, near Kula vill., Vidin region, N43°52'06'' E22°27'0'', 328m, 04.v.2010, 1 ♀ at light, leg. S. Beshkov (FBBZ). **New for Bulgaria.**

Distribution: Known from Central Europe, Romania and Greece (Razowski, 2002).

Cochylimorpha halophilana halophilana (Christoph, 1872) (Plate 2, 1, 2)

Bulgaria: Northern Black Sea Coast, Durankulak Lake near Shabla, N43°39'48'' E28°32'09'', 3 m, 09.x.2009, 1 ♂ at light, leg. B. Zlatkov, S. Beshkov; near Kranevo, N43°20'03'' E28°03'38'', 175 m, 22.viii.2010, 1 ♂ leg. B. Zlatkov & O. Sivilov; near Balchik, N43°23'56'' E28°10'53'', 35 m, 12.ix.2010, 2 ♀♀ at light, leg. B. Zlatkov & S. Beshkov; 15.ix.2010, 1 ♀ at light, leg. B. Zlatkov, S. Beshkov & C. W. Plant (FBBZ). **New for Bulgaria.**

Distribution: From Slovakia to South-Eastern Europe, Caucasus, Afghanistan (Razowski, 2002).

The specimens express wing pattern typical for ssp. *halophilana*. The colouration is the only reliable feature distinguishing the subspecies (Huemer, 2000). The genitalia do not show considerable differences. These of our specimens (male and female) are presented on Figs 1 and 3. The vesica everted (Fig. 2) is illustrated for the first time. Here we provide description of this important structure:

Length of phallus 0,94 mm, length of vesica 0,83 mm. The phallus (= aedeagus) is s-like bent, with sharp process at the apex. The vesica has tubular proximal part and widened distal part with one fingerlike diverticulum bearing fixed cornutus with socketed base and two pocket-like extensions. The surface of the diverticulum has circular rows of tiny spines visible only through compound microscope. A weakly sclerotised plate near the base of the diverticulum is present. The gonopore is undefined but it should be located between the

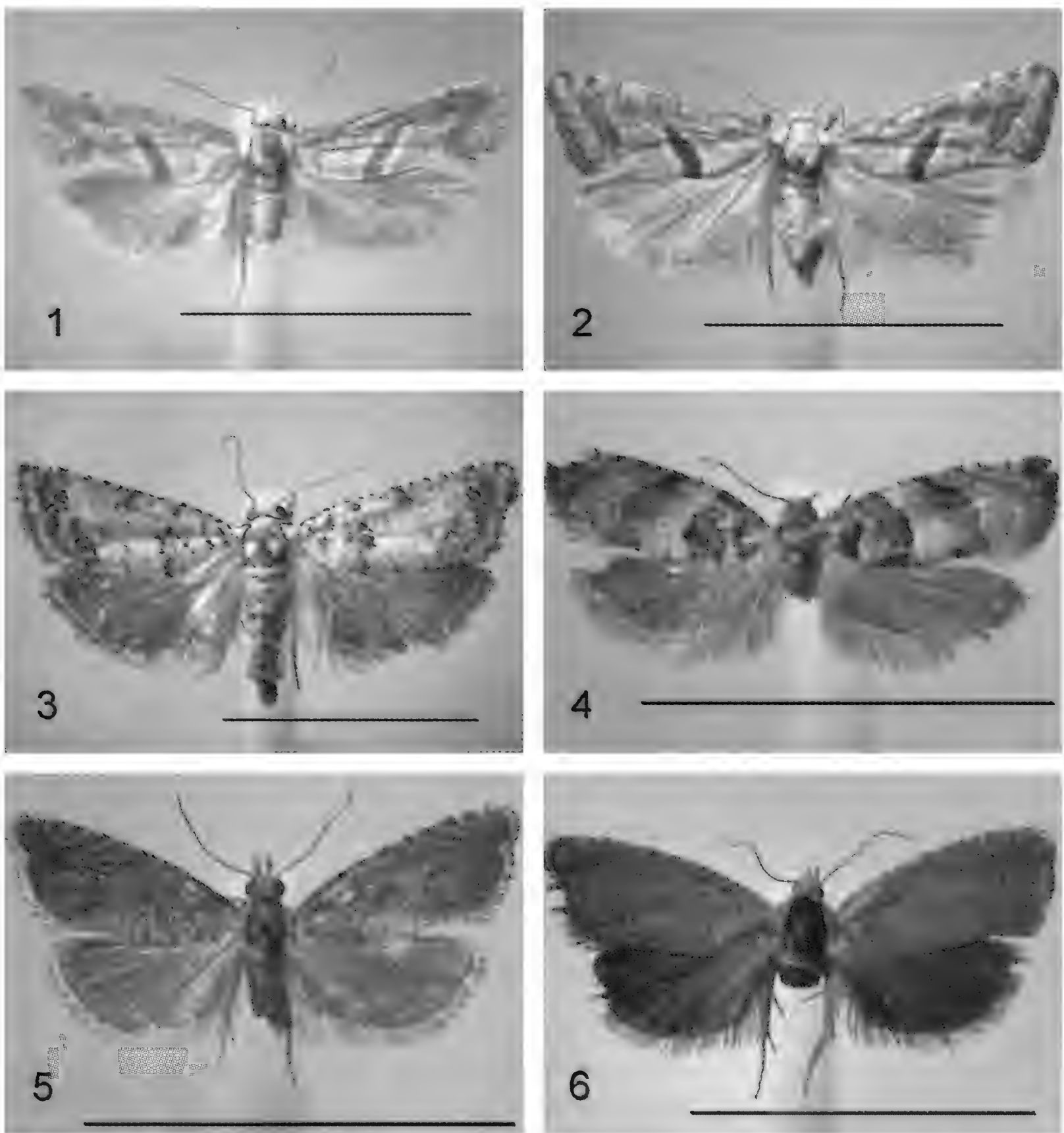


Plate 2. Tortricidae new for Bulgaria. 1, 2. *Cochylimorpha halophilana halophilana*: 1. ♂, Black Sea Coast, near Kranevo, 22.viii.2010; 2. ♀, near Balchik, 12.ix.2010. 3. *Epinotia dalmatana* ♀, near Novo Hodzhovo vill., 20.vii.2010. 4. *Gypsonoma obraztsovi* ♂, below Klyuch vill., 12.vi.2008. 5. *Dichrorampha caucasica* ♂, volcanic hill of Kozhuh, 24.x.2008. 6. *Dichrorampha baixerasana* ♀, University Botanical Garden in Varna, 05.vi.2010. Scale bar = 10 mm.

extensions. The main characters of the vesica are typical for tribe Cochylini: voluminous vesica with well developed diverticula and lateral gonopore.

All specimens were collected from a very limited area near the coast line (so called “Silver Coast”) consisting of white calcareous sediments (Plate 3). Unfortunately, the habitat (which is the only Bulgarian locality of several other moths, including the endemic *Oncocnemis michaelorum* Beshkov, 1997) is critically endangered by development for holiday homes and golf courses.

Aethes kindermanniana (Treitschke, 1830)

Bulgaria: SW Bulgaria, Kresna Gorge, N41°47'41" E23°09'39", 260 m, 14.viii.2006, 7 ♂♂, 2 ♀♀, leg. B. Zlatkov; 28.viii.2007, 2 ♂♂, 1 ♀ at light, 1 ♂ by netting, leg. B. Zlatkov & D. Gradinarov (FBBZ). **New for Bulgaria.**

Distribution: Europe excl. British Islands, Ural Mts.

Most specimens were collected flying around *Artemisia campestris* L. at twilight. *A. campestris*, which obviously is the food plant of the moth, is widely distributed in the gorge.

Cochylidia moguntiana (Rössler, 1864)

Bulgaria: SW Bulgaria, Kresna Gorge, N41°47'41" E23°09'39", 260 m, 04.vi.2006, 1 ♂, leg. B. Zlatkov; 24.iv.2007, 1 ♂, leg. B. Zlatkov & O. Sivilov; Petrich region, below Yavornitsa vill., N41°21'80" E23°01'50", 400 m, 20.iv.2007, 1 ♀, leg. B. Zlatkov & O. Sivilov (FBBZ). **New for Bulgaria.**

Distribution: Europe.

Cochylidia implicitana (Wocke, 1856)

Bulgaria: SW Bulgaria, Petrich region, below Klyuch vill., N41°23'05" E23°01'52", 225 m, 20.ix.2007, 2 ♂♂ at light, leg. B. Zlatkov & D. Chobanov; 30.iv.2008, 2 ♂♂ at light, leg. B. Zlatkov & D. Gradinarov; 12.vi.2008, 2 ♂♂, 1 ♀ in light trap, 1 ♂ by netting, leg. B. Zlatkov & O. Sivilov (FBBZ). **New for Bulgaria.**

Distribution: From Western Europe and North-Western Africa to Ural Mts., Siberia, Kazakhstan and Kyrgyzstan.

The specimens were collected from a wet habitat near Strumeshnitsa River.

Lobesia abscisana (Doubleday, 1849)

Bulgaria: SW Bulgaria, Petrich region, below Klyuch vill., N41°23'05" E23°01'52", 225 m, 12.vi.2008, 1 ♂ at light, leg. B. Zlatkov & O. Sivilov; marsh near Novo Hodzhovo vill., N41°24'26" E23°24'25", 115 m, 14.vi.2010, 1 ♀, leg. B. Zlatkov & O. Sivilov; W Bulgaria, Dragoman marsh, N42°56'13" E22°57'07", 706 m, 19.viii.2009, 1 ♂ at light, leg. B. Zlatkov, S. Beshkov & C. W. Plant (FBBZ). **New for Bulgaria.**

Distribution: Europe, Asia Minor, Transcaucasia, Western Siberia, Kyrgyzstan and Tajikistan.

All localities of this species in Bulgaria are marshy areas, although the food plants (*Anchusa*, *Echium*, *Artemisia* etc.) are widely distributed throughout the country.

Syricoris doubledayana Barrett, 1872

Bulgaria: SW Bulgaria, Petrich region, below Klyuch vill., N41°23'05" E23°01'52", 225 m, 12.vi.2008, 2 ♂♂ in light trap, leg. B. Zlatkov & O. Sivilov; Struma valley, marsh near Novo Delchevo, N41°30'06" E23°17'51", 140 m, 23.v.2010, 3 ♂♂ in light trap, leg. B. Zlatkov, O. Sivilov & G. Hristov (FBBZ). **New for Bulgaria.**

Distribution: widely distributed throughout Eurasia.

This species seems to be relatively rare in Bulgaria. It was found in marshy meadows near Petrich with hygrophilous vegetation: *Phragmites*, *Typha*, *Sparganium*, *Cyperus*. The last one is believed to be one of the food plants of the larva (Razowski, 2003).

Epinotia dalmatana (Rebel, 1891) (Plate 2, 3)

Bulgaria: SW Bulgaria, Kresna Gorge, Stara Kresna railway station, 30.vii.1975, 1 ♂, leg. A. Slivov (IBER); near Novo Hodzhovo vill., N 41°24'27" E 23°25'01", 200 m, 20.vii.2010, 1 ♀ in light trap, leg. B. Zlatkov & O. Sivilov (FBBZ). **New for Bulgaria.**

Distribution: Southern Europe: Spain and France to Dalmatia, southern part of Eastern Europe; Asia Minor, Iran, Iraq, Syria, Transcaucasus (Razowski, 2003).

Both localities of this Mediterranean species are dry, hot, bushy habitats.

Gypsonoma obraztsovi Amsel, 1959

Bulgaria: SW Bulgaria, Petrich region, below Klyuch vill., N 41°23'05" E 23°01'52", 225 m, 12.vi.2008, 1 ♂ at light, leg. B. Zlatkov & O. Sivilov (FBBZ). **New for Bulgaria.**

Distribution: Hungary (<http://www.faunaeur.org>), Romania, Asia Minor, Israel, Iran, Caucasus, Transcaucasus, Tajikistan (Razowski, 2003).

Only the males of this rare species are known. The specimen collected shows darker ground colour of the forewings (Plate 2, 4) in comparison with the illustrated by Razowski (2003, Pl. XI, Fig. 314). Genitalia are typical, however (Fig. 4).

Epiblema cnicicolana (Zeller, 1847)

Bulgaria: SW Bulgaria, Struma River Valley, volcanic hill of Kozhuh, N 41°27'43" E 23°15'30", 200 m, 23.v.2010, 1 ♂, leg. B. Zlatkov, O. Sivilov & G. Hristov (FBBZ). **New for Bulgaria.**

Distribution: known from many European countries: from England to Germany, Slovakia to Romania, Southern Europe from Spain to Sicily and Albania (Razowski, 2003).

Corticivora piniana (Herrich-Schäffer, 1851)

Bulgaria: SW Bulgaria, the western foothills of Pirin Mts., Ilindentsi vill., N41°39'05" E23°14'47", 475 m, 14.vi.2008, 1 ♀, leg. B. Zlatkov & O. Sivilov (FBBZ). **New for Bulgaria.**

Distribution: Almost entire Europe.

The specimen was collected in a small pine plantation during late afternoon.

Cydia conicolana (Heylaerts, 1874)

Bulgaria: SW Bulgaria, Slavyanka Mts., the road to Livade place, N41°24'57" E23°36'51", 1550 m, 15.vi.2010, 1 ♂ at light, leg. B. Zlatkov & O. Sivilov (FBBZ). **New for Bulgaria.**

Distribution: from the British Islands and Netherlands to Greece, Asia Minor and Caucasus (Razowski, 2003).



Plate 3. Endangered habitat of *Cochylimorpha halophilana halophilana* near Balchik, 19.viii.2009.

Cydia cosmophorana (Treitschke, 1835)

Bulgaria: SW Bulgaria, Pirin Mts., above Sugarevo vill., N 41°34'19" E 23°26'12", 885 m, 17.vi.2010, 1 ♀ at light, leg. B. Zlatkov & O. Sivilov (FBBZ).

New for Bulgaria.

Distribution: Europe through Siberia to Russian Far East (Razowski, 2003, <http://www.faunaeur.org>).

Pammene luedersiana (Sorhagen, 1885)

Bulgaria: SW Bulgaria, Struma valley, Dolno Gradishte place near Kamenitsa vill., N 41°38'22" E 23°09'54", 250 m, 16.iv.2006, 1 ♀, leg. B. Zlatkov; 05.iv.2009, 1 ♂, 2 ♀♀, leg. B. Zlatkov; Rupite, near Petrich, Kozhuh hill, N 41°27'43" E 23°15'30", 200 m, 15.iv.2006, 1 ♀, leg. B. Zlatkov. All specimens were collected by net at late afternoon.

Distribution: Europe, Asia Minor, Caucasus.

The specimens were collected from two hot and dry habitats in SW Bulgaria. This fact contradicts our speculation that the species could be found in Bulgaria at higher altitude. According to some authors (Danilevsky & Kuznetzov, 1968; Razowski, 2003) the moth occurs in peat-bogs and the food plant of the larvae is *Myrica gale*. This plant has never been found in Bulgaria and we suppose that *P. luedersiana* feeds also on other species.

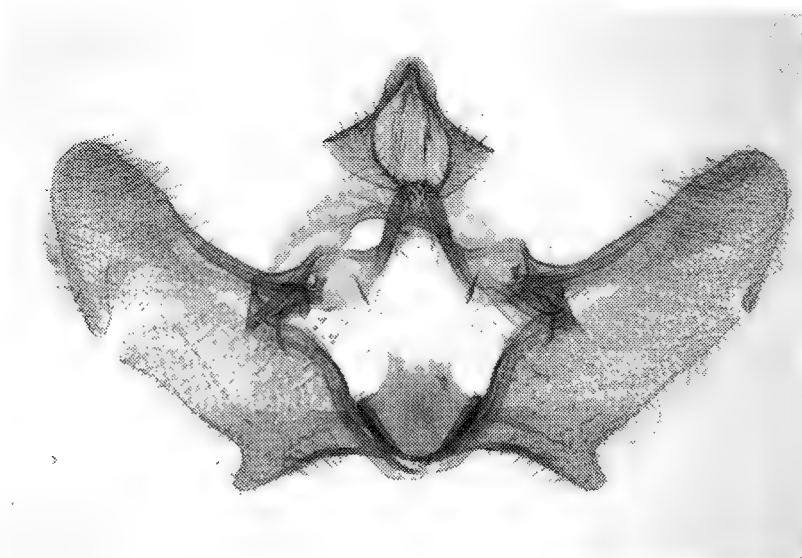


Fig. 1. *Cochylimorpha halophilana*, male genitalia (without aedeagus), Black Sea Coast, near Kranevo, 22.viii.2010.

Fig. 3. *Cochylimorpha halophilana*, female genitalia, Black Sea Coast, near Balchik, 12.ix.2010.

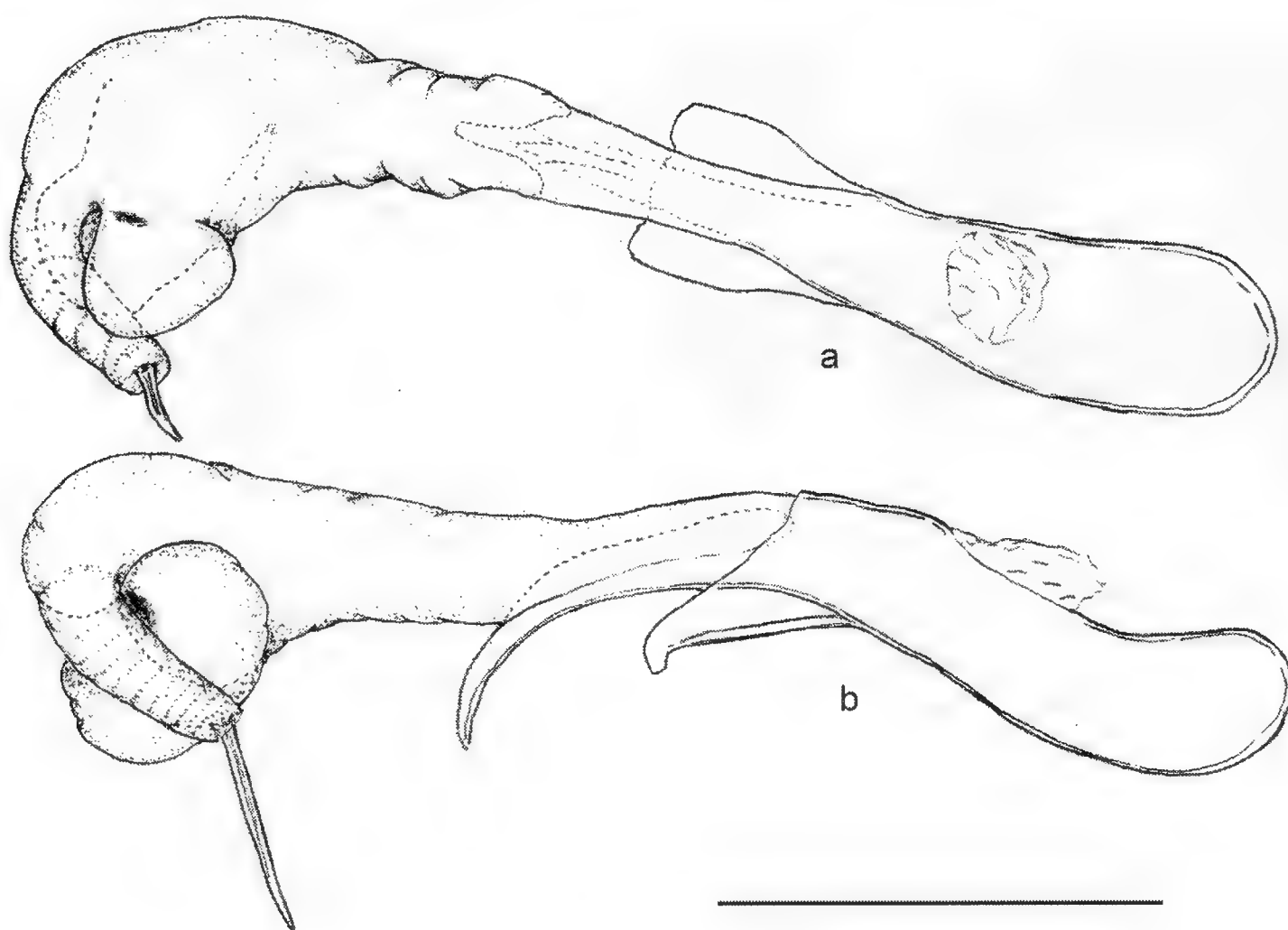


Fig. 2. Everted vesica of *Cochylimorpha halophilana halophilana*, the specimen from Fig. 1. a, dorsal view; b, lateral view. Scale bar = 0,5 mm.

During dissection of the male genitalia the vesica was successfully everted (Fig. 5), therefore we provide a description and illustration of this structure for the first time. Length of phallus 0,54 mm, length of vesica ~ 0,29 mm. The phallus is tubular with wide base and narrower distal part; coecum absent. The vesica is typical for Grapholitini: with cylindrical basal section and bulbous, dorsally oriented rather voluminous distal section, distinctly separated from the ductus ejaculatorius. The bulbous part bears two groups of ventrally located cornuti. The proximal group consists of 13 fixed cylindrical spiniform cornuti with sockets, and the distal one has three lanceolate deciduous cornuti with lateral bases.

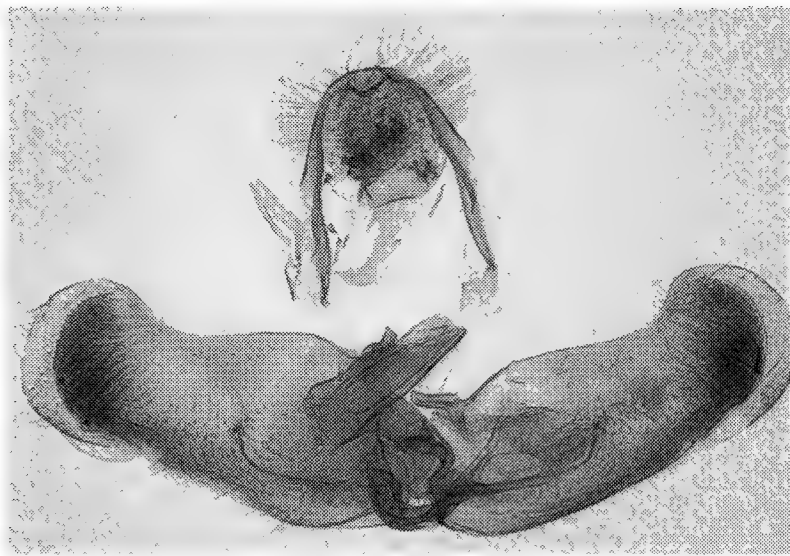


Fig. 4. *Gypsonoma obraztsovi*, male genitalia, below Klyuch vill., 12.vi.2008.

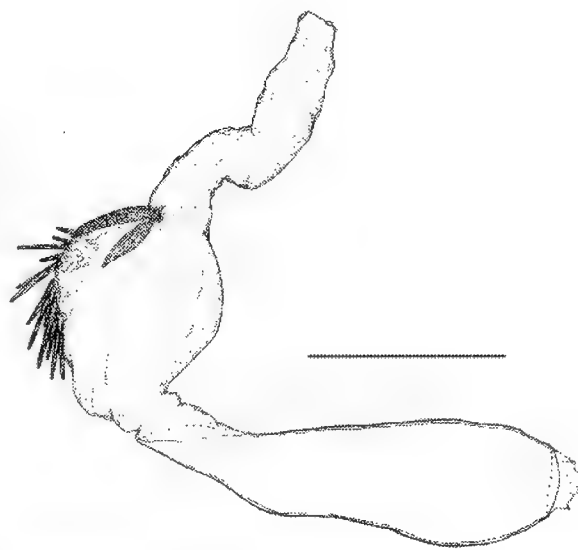


Fig. 5. Everted vesica of *Pammene luedersiana*, Struma valley, Dolno Gradishte, near Kamenitsa vill., 05.iv.2009. Scale bar = 0,25 mm

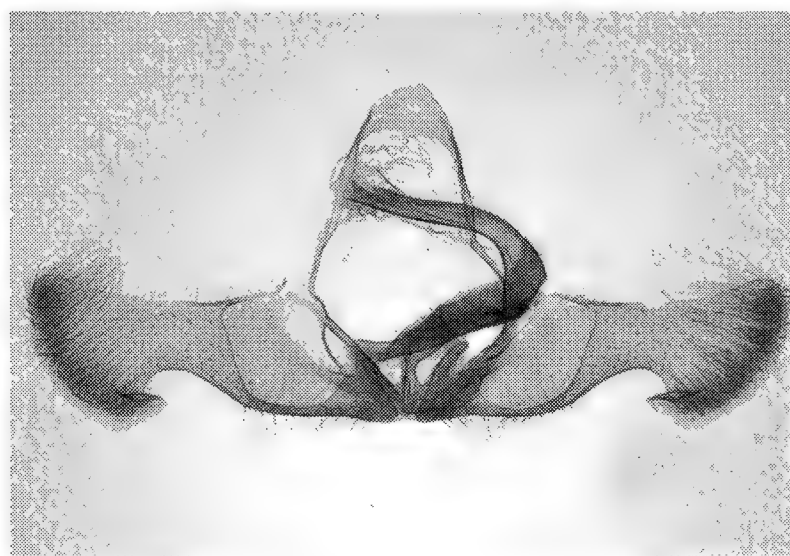


Fig. 6. *Dichrorampha caucasica*, male genitalia, volcanic hill of Kozhuh, 24.x.2008.



Fig. 7. *Dichrorampha baixerasana*, female genitalia, University Botanical Garden in Varna, 05.vi.2010.

Gonopore is located apically. An interesting fact is that the vesica bears two types of socketed cornuti: deciduous and non-deciduous.

Dichrorampha caucasica (Danilevsky, 1948)

Bulgaria: SW Bulgaria, Kresna Gorge above Oshtavska River, N 41°45'35" E 23°09'30", 290 m, 11.x.2011, 1 ♂, at light, leg. B. Zlatkov & O. Sivilov; Struma River Valley, volcanic hill of Kozhuh, N 41°27'43" E 23°15'30", 200 m, 24.x.2008, 2 ♂♂, at light, in early morning (~ 7 h a.m.), leg. B. Zlatkov & D. Gradinarov; 13.x.2011, 1 ♂ in light trap, leg. B. Zlatkov & O. Sivilov. **New for Bulgaria and the Balkan Peninsula.**

Distribution: Ukraine: Crimea (Budashkin, 2009), Caucasus, Transcaucasia, Kopet Dag, Asia Minor (Kuznetzov, 1978).

The specimens from 2008 were collected in the early morning, just before sunrise, at "light tower". The weather was windy in the evening, but calm in the morning, when the specimens were attracted. The specimen from Kresna Gorge was collected in the evening, i.e. this species should have two periods of activity.

A closely related species, *Dichrorampha coniana* Obraztsov, 1953, is reported for Europe from Macedonia (Razowski, 2003). Danilevsky & Kuznetzov (1968) believe that *D. coniana* should be a junior synonym of *D. caucasica*, but they do not synonymise it. According to these authors the specimen described by Obraztsov (1953) as *coniana* may be a local form of *caucasica*. The illustration of Obraztsov (*op.cit.*, Fig. 16 on p. 42) of *coniana* is rather schematic, and the differences with *caucasica* may be due to deformation by the cover glass. Both species have very similar male genitalia (compare Pl. 51, Fig. 520 in Razowski, 2003, and Fig. 80a, p. 176 in Danilevsky & Kuznetzov, 1968). Small differences can be seen in the lower edge of the cucullus, but the aedeagus is identical and we consider the opinion of Danilevsky & Kuznetzov (*op. cit.*) as correct. We did not see the specimens from Macedonia and can not comment them, but the Bulgarian specimens fit very well the genitalia figured by Danilevsky & Kuznetzov (*op. cit.*) (Fig. 6), and we believe that they are *D. caucasica*. It is more likely if the Macedonian material is also *D. caucasica*. A Bulgarian specimen of this species is shown on Plate 2, 5. It is interesting to note that *D. caucasica* is a common species in Caucasus at high altitude (1600-1800 m). The Bulgarian localities are much lower, 200-500 m alt.

Dichrorampha baixerasana Trematerra, 1991

Bulgaria: Black Sea Coast, University Botanical Garden in Varna, N 43°14'04" E 28°00'02", 05.vi.2010, 1 ♀, leg. B. Zlatkov at late afternoon.

Distribution: Italy, Croatia, Albania (Razowski, 2003).

This species is locally distributed in the Mediterranean belt of Europe. Its founding on the Bulgarian Black Sea Coast is interesting but not surprising. Externally *D. baixerasana* is similar to some other *Dichrorampha* species as *senectana* Guenée, *obscuratana* (Wolff, 1995) and even *cinerosana* (Herrich-Schäffer, 1851). The female genitalia (Fig. 7) are similar to these of *D. senectana*, but the colliculum is distinctly longer. The specimen collected is well preserved, with very dark hind wings (Plate 2, 6).

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The Isle of White Wave *Idaea humiliata* (Hufn.) (Lep.: Geometridae) found in Old Weston, Huntingdonshire

On the evening of 4 July 2011, KR put his 125-watt mercury vapour moth trap in his garden as usual. The following morning when emptying the trap an unknown geometrid was found. After searching through the literature he was still not sure what the moth was, so put it aside for genitalia examination later in the year.

On 23 October 2011, KR dissected the moth and came up with a preliminary identification, which he thought impossible. He then e-mailed photographs of the genitalia, aedeagus and the moth to BD with the following comment “The attached is causing me some concern. It is one of the Waves but the nearest I can get seems impossible. I’ll tell you my thought after you tell me what it is.” BD examined the photographs and after referring to Hausmann (2004. Sterrhinae. – *In* A. Hausmann (ed.): *The Geometrid Moths of Europe* 2: 1-600), replied “I suspect you are thinking of Isle of Wight Wave. (Am I correct?). This is the closest I can get. However, it is not impossible as it is still common on the continent and from

the European maps I have it looks like it is very common in Holland with scattered records in the north of France. We have had a lot of southerly winds lately.”

BD then decided to send the photographs to John Langmaid who also agreed that it was most probably Isle of Wight Wave and suggested it was sent for a further opinion. This BD did, but failed to receive a reply despite a reminder being sent a couple of weeks later. BD then decided to ask Jon Clifton his opinion. Jon replied and agreed with the identification and said that he had one in his collection from France.

The Isle of Wight Wave is reported as being extinct in Britain, but was formally a resident on the Isle of Wight, but has not been seen there since 1931. A single specimen was trapped in Portsmouth, Hampshire in 1954.

We would like to thank John Langmaid and Jon Clifton for their help with the identification of this moth. — KEVIN ROYLES, Brook Farm House, Main Street, Old Weston, Huntingdon PE28 5LL (E-mail: janeroyles@btinternet.com) and BARRY DICKERSON, 27 Andrew Road, Eynesbury, St Neots PE19 2QE (E-mail: barry@eynesbury27.freeseve.co.uk).

***Ponera testacea* (Emery) (Hym.: Formicidae) in Devon and Cornwall**

Attewell et al (2010. *Ent. Rec.* **122**: 113-119) drew attention to the discovery of the ant *Ponera testacea*, new to Britain, from Dungeness, East Kent. This species had been regarded as a variety of *P. coarctata* (Latreille) until raised to species level by Csoz & Seifert (2003. *Acta Zoologica Academiae Scientiarum Hungaricae* **49** (3): 211-233). Paul (*BWARS Newsletter Autumn 2011*: 24-25) subsequently re-examined the five voucher specimens from West Sussex and the Isle of Wight in his collection and found them all to be *P. testacea*. Following his lead, the author re-examined his own 8 *Ponera* vouchers – all workers – and these too are all *P. testacea*. The data are:

East Cornwall (VC2): Pentire Head (SW9280), one beneath thyme mat on outcropping rock on south-facing sea cliff, 2.viii.2006. This record was published as *P. coarctata* (Alexander, 2007. *Br. J. Ent. Nat. Hist.* **20**: 141). **S. Devon** (VC3): Slapton Bar (SX8243), one from sparsely-vegetated coastal shingle bar, 4.ix.2003; Copstone Cove, Chivelstone (SX7735) 31.vii.2007; Hilsea Point, Newton & Noss (SX5445) 24.vii.2007 (3 specimens); Stoke Point (SX5645) 19.vii.2007; and Turnchapel Quarry, Plymouth (SX4952) 7.vi.2011.

The South Devon ants were all taken by suction sampling using a domestic two-stroke leaf sucker/blower machine, while the Cornish specimen was found by hand-searching. These records bring the known distribution of this ant right across the English south coast from Kent to Cornwall.

It would seem likely that this is a long-established native species, although more re-examination of voucher collections is needed to confirm this. *P. testacea* appears to be associated with xerothermic, sparsely-vegetated situations, apparently avoiding more humid sites such as open xerothermic woodland. In contrast, the true *P. coarctata* favours warmth and humidity, tolerating partial shade. *Ponera* have been known from the south Devon coast for many years and it seems reasonable to assume that all of these records refer to what we now call *P. testacea*, at least until more vouchers are checked.

My thanks to Mike Fox and Phil Attewell for their interest in my records.—
KEITH N. A. ALEXANDER, 59 Sweetbrier Lane, Heavitree, Exeter EX1 3AQ (E-mail: keith.alexander@waitrose.com).

ANNOUNCEMENT: GELECHIID RECORDING SCHEME

A new Gelechiid Recording Scheme was launched in January 2012 with a supporting website, still under development, hosted by the Biological Records Centre based at CEH Wallingford. **The aims of the Scheme are to:**

- encourage interest in, and the recording of Gelechiid moths (Lepidoptera: Gelechiidae) occurring in the British Isles (including the Channel Islands)
- produce readily accessible species distribution maps at a 10 km map square level
- improve knowledge of the status of all members of this family to aid their conservation

A total of 163 different species of gelechiid moths have been recorded in Great Britain and Ireland although several are now considered extinct while a few are accidental arrivals on imported goods. As data become available maps will be placed on the scheme website at www.gelechiid.co.uk.

Records of all Gelechiid species recorded in the British Isles and Channel Islands are welcomed and should be sent by e-mail, or to the postal address at the end of this article. **Ideally** the information should include:

Species*	full binomial (plus Bradley and Fletcher number if known)
Site name	
O.S. grid reference*	Ideally to 6 figures, though four-figures will do and two are acceptable if that is all that is available
Vice County Number	If known
Recorder*	
Identifier	
Date*	As precisely as possible. Please record overnight sessions as the earlier date (e.g., 14 – 15 March should be recorded as 14 March not 15 March)
Quantity	Where known
Sex	If known (e.g., if dissected)
Method of recording	e.g. light trap, bred ex larva
Stage*	e.g. adult, larva
Comments	e.g. genitalia dissection, foodplant if bred

* minimum requirements.

MapMate files, Excel files, MS Word documents or even hand written lists are acceptable, but please, if at all possible, avoid sending records in tabular form with species down one side and dates along the top. If you have data in a different format then do please get in touch to discuss the options. To enable data entry to be as rapid as possible, MapMate users are requested to provide an import ready text file (please get in touch for information on how to carry this out) – it only takes a couple of minutes to create. It will *not* be possible to accept Mapmate Sync files.

MapMate Ltd have kindly produced a patch to enable quick and easy creation of a data file covering this scheme (issued in Patch number 431 issued on 24 July 2011). It overwrites a previous patch produced for the 2001 “Gelechiid Plus Scheme”, which should not be used. Annual updates of new and edited data would be appreciated as convenient.

If we can be of any assistance in carrying out dissections do please get in touch. We look forward to hearing from you and please don't hesitate to contact us if you have any questions or concerns. Please note that, unless requested otherwise, any records contributed to the Gelechiid Recording Scheme will be shared with other moth recorders, Butterfly Conservation, County Moth Recorders, the National Biodiversity Network (NBN) and with other conservation organisations as appropriate.— CAROLYN and STEPHEN PALMER, 137 Lightfoot Lane, Fulwood, Preston, Lancashire PR4 0AH (E-mail: s.palmer12@btopenworld.com).

BOOK REVIEWS

Distribution Atlas of Butterflies in Europe by Otakar Kudrna, Alexander Harpke, Kristian Lux, Josef Pennerstorfer, Oliver Schweiger, Josef Settele and Martin Wiemers. 576pp., including full colour distribution maps of 441 butterfly species in Europe and some colour photographs. 175 x 245 mm., hardbound, ISBN: 978-3-938249-70-3. Published by Gesellschaft für Schmetterlingsschutz e.v. (Society for the Conservation of Butterflies), Topasweg 12, D-06120 Halle, Germany (meb-gfs@ufz.de), 2011. 65 Euros (£60). Available from Pemberley Books (www.pemberleybooks.com) and other natural history book sellers.



It is nine years since Otakar Kudrna's *The Distribution Atlas of European Butterflies* was published in March 2002, the first such atlas to cover the butterflies of a whole continent. This successor is much more than a new edition and has been entitled *Distribution Atlas of Butterflies in Europe*. Again spear-headed by Otakar Kudrna, a well known and distinguished Czech entomologist based in Germany, he has been assisted this time by Alexander Harpke, Kristian Lux, Josef Pennastorfer, Oliver Schweiger, Joseph Setterle and Martin Wiemers. They have assessed some 655,000 data records, nearly three times the total used for the first atlas, to produce full colour distribution maps for all the 441 European species of butterflies. As stated in the summary on the back

cover of the book, apart from providing comprehensive chorological data for biogeographers and taxonomists, the prime purpose of this atlas is to aid and promote the conservation of European butterflies. This is especially facilitated by the provision of coincidence maps showing the distribution of species diversity and thus revealing those areas with the highest density of species; the distribution and density of endemic species, particularly rare endemics and those species on the European Red List – thus highlighting those areas with the highest totals of such species and also those that are the most threatened. These are indicated by scales shading from pale pink for the lowest categories to deep red for the highest. Although they convey a good general impression of degree on the maps, it is difficult to distinguish between very similar shades. Nevertheless, these maps, together with the species maps, do clearly identify those areas of significant conservation importance to which priority should be given.

In the Preface, the senior editor and instigator of the whole mapping project, Dr Kudrna, now retired, sets forth his reasons for taking on the task of producing this atlas and its predecessor. He realised that the then existing 'sub-matchbox-sized' and often inaccurate distribution maps in the field guides of the 1970s were quite inadequate for many purposes and needed to be improved upon. In particular, he considered that, in view of the rapid decline of many European butterfly species, priority objectives for their conservation could not be readily ascertained without a comprehensive distribution atlas. He therefore set up the Mapping European Butterflies (MEB) project in 1996, dedicating much hard work to the task, and enlisting the help of 254 contributing recorders for the first atlas (MEB1) published in 2002 and 272 for the present one (MEB2).

With the help of his co-workers, especially Josef Setterle, Kudrna has completely revised and rewritten the 2002 atlas. He had hoped for funding support for the publication from the European Union, but this was not forthcoming. However, from 1995 the project was backed by the Naturschutzbund Deutschland (NABU) and the Gesellschaft für Schmetterlingsschutz (GfS) with the objectives of setting-up a distribution data bank of European butterflies; the publication of a European distribution atlas containing dot maps; the assessment of the conservation status of the European butterfly species; and the identification of priorities and high priority areas for their conservation.

The standard maps in the atlas show the area of Europe covered: all western Europe east as far as the eastern frontiers of Norway, Finland, Estonia, Latvia, Belarus, the Ukraine, Rumania, Bulgaria and Greece, and including European Turkey, the Greek islands, Cyprus and other European islands in the Mediterranean and the Atlantic.

A reference locality system (RL), based upon the *Times Atlas of the World*, was chosen for the area covered by each dot on the map. Red dots were used for the period before and including 31 December 1950; yellow dots for that between 1 January 1951 and 31 December 1980; and green dots for that after and including 1 January 1981. Each RL could cover up to 50 square kilometres. The quantity and quality of the coverage varied from generally very good in western European countries, such as the United Kingdom, Germany and the Netherlands, to poor in some eastern countries, such as Albania, Belarus and Moldova. No data was received from the Russian enclave in the former East Prussia. A lot of the editors' time was spent in correcting errors in the data received, often resulting from recorders confusing latitude with longitude! Recorders and supporters of massive project are acknowledged in full on pages 16-21.

In introducing two checklists of species included in the Atlas, one arranged systematically, the other alphabetically, Kudrna criticises the use of vernacular names instead of valid scientific names by some conservationists and ecologists, and also the alternative practice of using the vernacular as the 'primary' name and the scientific name as a 'second rate appendix'. However, he accepts that the 'occasional use of vernacular names supplementing scientific names of common species easily recognizable to laymen in the language of the country concerned, may be acceptable in publications intended primarily for widespread general use.' Scientific names only are used throughout the Atlas and reveal Kudrna's 'lumper' preference, although some compromises have been made, such as in the use of *Plebejus* and *Aricia* for some of the Polyommatae. Some of the names chosen may be unfamiliar to British entomologists (e.g., *Ochlodes sylvanus*). The systematic checklist included in the Atlas has been revised and updated, but an urgent demand is nevertheless made for a standard checklist. In addition to the two checklists, a synonymic list of species names often used in field guides is also included.

The bulk of the Atlas (pp.49-483) consists, of course, of well presented colour distribution maps (prepared by Alexander Harpke) for all 441 breeding European species. These measure 135mm x 135mm and are on a scale of 1:37.000.000 and are arranged alphabetically according to generic names. The caption for each map contains basic species-specific information, where relevant, on zoogeography, conservation and taxonomy. Under the heading of Zoogeography, basic simplified information on the world range of the species is given, while under Conservation the relationship between the European distribution of the species and its world range is expressed, its status according to the IUCN's European Red List, its status under the EU's 'Habitats' Directive, and an assessment of the risk to which it is exposed as a result of the changing climate. The notes under the Taxonomy heading provide supplementary information on the taxonomic status, identity, nomenclature, affinity or any other species relevant aspect that may be necessary or useful. They also give

explanations for and justify the reasons for the authors' taxonomic judgments and conclusions.

Following the maps there are sections discussing the analysis and evaluation of the data on which they are based. The coverage obtained is considered to be extremely satisfactory (95% of the basic recording fields (BRFs)). Doubtful records were rigorously excluded. A summary of the coverage and state of recording is given for every European country. Under the heading Zoogeography (pp. 489-497) there is a discussion of the faunal elements (defined as 'an assemblage of species showing similar present ranges') to which the various European butterfly species belong. Coincidence maps show the concentrations of butterfly diversity ('hot spots'), endemic species and those threatened with possible extinction, plus those protected by the EU FFH-Directives.

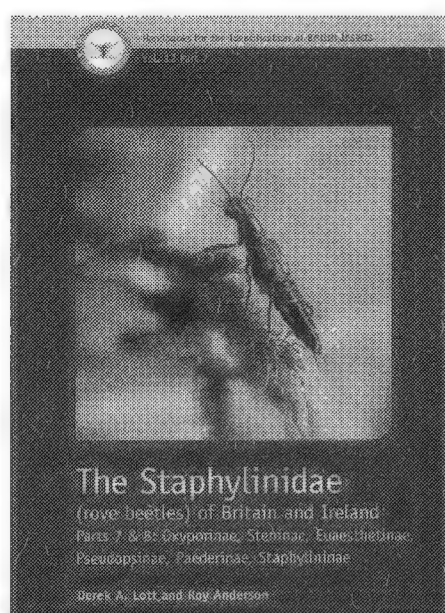
The Atlas concludes with a discussion of the outlook for the successful continuance of butterfly mapping in Europe, a sample standard MEB Recording and Monitoring Sheet, an extensive bibliography, a gazetteer of reference localities, a list of abbreviations, acronyms and definitions, and an index of the scientific names of the species. An appeal is made in the Outlook section (p.505) for new recorders to fill existing gaps, especially in those areas and countries where present coverage is poor.

In the opinion of this reviewer, the Atlas fully lives up to the claims made for it. It enables one to see at a glance a clear and accurate cartographical portrayal of the European distribution of each species so far as current information is available, and will be a valuable tool in assessing the priorities for the conservation of butterflies, important indicators of environmental quality. This reviewer has noticed very few typographical errors in the text, which has been written in generally straightforward, clear English. The references on pp. 489-491 to an important paper by R. H. L. Dennis, stated there to have published in 1995, was actually published in 1998 and has been accidentally omitted from the bibliography.

Despite these minor points, the editors, Dr. Kudrna in particular, are to be congratulated on producing a most useful and valuable book that should be on the bookshelves of every Lepidopterist seriously interested in European butterflies, and every wildlife conservationist and, I suggest, climatologists as well.

John F. Burton

The Staphylinidae (rove beetles) of Britain and Ireland Parts 7 & 8: Oxyporinae, Steninae, Euaesthetinae, Pseudopsinae, Paederinae, Staphylininae by Derek A. Lott and Roy Anderson. Handbooks for the Identification of British Insects Vol. 12 Parts 7-8, published for the Royal



Entomological Society by the Field Studies Council, 2011. Colour plates by James Turner, edited by Mike Wilson. 340 pp, 190 colour plates, 429 figures. Royal Entomological Society, The Mansion House, Bonehill, Chiswell Green Lane, Chiswell Green, St Albans, AL2 3NS. ISBN 978 0 901546 92 0. Softcover. Price £49.00.

Rove beetles have gained the reputation in Coleoptera circles of being a 'difficult' group and many beginners are reluctant to tackle this large family. This is not too surprising because, until very recently, the only works in English where a student stood any chance of arriving at the correct name were Joy's Practical

Handbook of British Beetles, published almost 80 years ago, and the Handbook by C. E. Tottenham, published by the Royal Entomological Society of London in 1954. Whilst both works proved very useful in the past there have been so many changes in recent years that the keys and descriptions they contain have become unreliable when used on their own. Alternative keys to most species occurring in the British Isles do exist, but these are mostly published in languages other than English and furthermore no single foreign work is likely to contain every species contained in our checklist.

The appearance of this excellent volume is a welcome addition to the RES Handbooks series and sets a new standard for others to follow. It is the second part in a series that will cover the whole of what are currently regarded as Staphylinidae and includes four of the subfamilies dealt with in Tottenham's handbook (Oxyporinae, Steninae, Euaesthetinae and Pseudopsinae) plus Paederinae and Staphylininae.

The book follows a similar format to other recently published RES Handbooks. The Contents page provides a quick reference to all the chapter headings and includes scientific names to genus level. This is followed by short sections entitled "Morphology", "Biology", "Collection methods and preservation", "Notes on keys", "Glossary" and "Classification and nomenclature". The last section includes a useful key to all subfamilies of Staphylinidae. A detailed checklist of species covered in this volume is followed by keys to species and a numbered paragraph dealing with each taxon in turn, including a description, notes on similar species, habitat and distribution.

In most cases the book avoids couplets that rely on comparative characters that all too frequently either cause confusion or prevent a positive determination when closely related species are not available for reference. The illustrations are of extremely high quality and include the aedeagus of almost every species. Many important characters are illustrated by drawings placed beside the couplets but genitalia drawings are generally grouped together on one page, although they are never very far removed from the relevant text. Almost two-thirds of the species are illustrated by colour photographs at the end of the book. Whilst these can be extremely helpful in pointing a student in the right direction, it is important to realise that most rove beetles cannot be determined by referring to photographs alone.

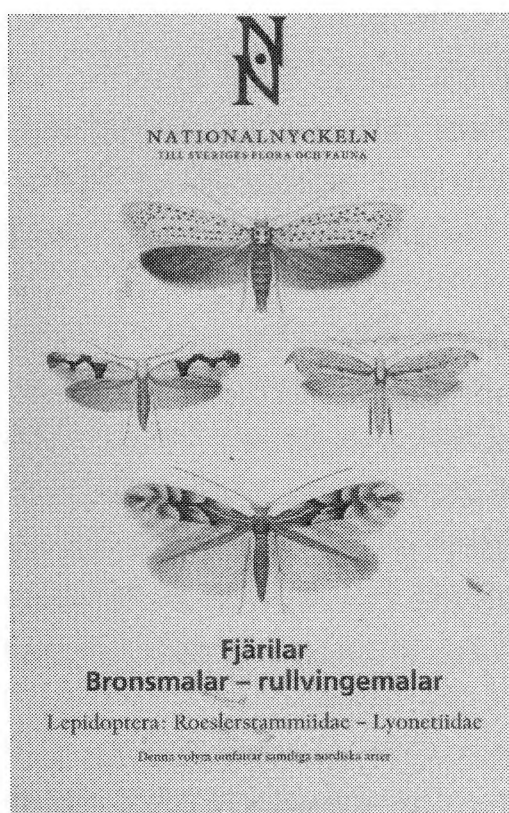
Several recent additions to the fauna of the British Isles are included, for example: *Edaphus beszedesi* Reitter, *Philonthus alpinus* Eppelsheim, *Astrapaeus ulmi* (Rossi), *Quedius lucidulus* Erichson, *Q. lyszkowskii* Lott and *Gyrohyphnus wagneri* (Sheerpeltz).

This handbook represents a huge step forward in the study of Staphylinidae and it is now possible to accurately identify almost half the recorded British and Irish species with a high degree of certainty by referring to this work and its companion volume [Lott, D. A. 2009. The Staphylinidae (rove beetles) of Britain and Ireland Part 5: Scaphidiinae, Piestinae, Oxytelinae. *Handbooks for the Identification of British Insects* Volume 12 Part 5].

This book is a credit to the authors who must be congratulated for producing such a thorough work that will undoubtedly become an important source of reference for many years to come. There is one misprint that might lead to confusion when quoting a reference. At the top of the front cover it states "Vol. 12 Part 7" but on the spine it reads "Parts 7 & 8). It is presumed that the latter is correct and the reference should be "Lott, D.A. & Anderson, R. 2011. The Staphylinidae (rove beetles) of Britain and Ireland, Parts 7 and 8: Oxyporinae, Steninae, Euaesthetinae, Pseudopsinae, Paederinae, Staphylininae. *Handbooks for the Identification of British Insects* 12(7-8): 1-340".

Peter Hodge

Nationalnyckeln till Sveriges Flora och Fauna. Fjärilar: Bronsmaler – rullvingemalar (Lepidoptera: Roeslerstammiidae – Lyonetiidae) by B.Å. Bengtsson & R. Johansson. 494pp., 230 x 288 mm., hardbound, ISBN: 978 91 88506 50 4. Published by ArtDatabanken SLU, Uppsala, 2011. Available from UK booksellers at £49 plus postage.



This book is the third volume on Lepidoptera in a series covering the flora and fauna of Sweden. It is lavishly produced and very reasonably priced since it is heavily subsidised by the Swedish Government. Since this series is about the Swedish fauna the text is in Swedish, but abstracts of all species are given in English.

The introductory pages explaining the systematic position of the Lepidoptera are solely in Swedish. There is then a list of species and a key to families. The sequence is very similar to that used in British lists, but in contrast with most European works there is very zealous adherence to the ICZN code so that many terminations are unfamiliar. For example the genus *Phyllonorycter* is deemed to be masculine, so that most species names, in order to agree with it, end in *-us* for the first time in any literature that I have seen. Other workers favour the use of original spelling

to avoid changes in ending every time a species is assigned to a different genus.

The systematic treatment of each species contains beautifully executed paintings of adults and line drawings of genitalia and wing venation. There is a distribution map covering Scandinavia. For each family a colour photograph of a live specimen is included. The families treated in this volume are: Roeslerstammiidae, Douglesiidae, Bucculatricidae, Gracillariidae, Yponomeutidae, Plutellidae, Acrolepiidae, Glyphipterigidae & Lyonetiidae.

After the descriptions there is a key to larvae and mines or other larval habitation, arranged in alphabetical order of plant genus. Next the illustrations of adults are repeated so that comparison of similar species can be made.

Most British species are treated in this book, but there are a number which are absent from our fauna, and likewise a few are omitted which do not occur in Sweden.

Photography has improved so much that generally photographs are preferable over paintings and drawings, but with very small moths it is often difficult to obtain a perfect specimen of each species. Roland Johansson's illustrations surpass anything I have seen and one has to admit they are at least as good as photographs. The English descriptions are concise but accurate.

Although the bulk of the book is in Swedish it will still be found very useful to British entomologists, and very good value. I commend it with enthusiasm.

David Agassiz

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